REVIEW



Coronavirus in human diseases: Mechanisms and advances in clinical treatment

Panpan Lin^{1,#} | Manni Wang^{1,#} | Yuquan Wei¹ | Taewan Kim² | Xiawei Wei¹ |

¹ Laboratory of Aging Research and Cancer Drug Target, State Key Laboratory of Biotherapy and Cancer Center, National Clinical Research Center for Geriatrics, West China Hospital, Sichuan University, Chengdu, China

² Wexner Medical Center, The Ohio State University, Columbus, Ohio 43210, USA

Correspondence

Taewan Kim, Wexner Medical Center, The Ohio State University, Columbus, OH 43210.

Email: kim.2203@buckeyemail.osu.edu Xiawei Wei, Laboratory of Aging Research and Cancer Drug Target, State Key Laboratory of Biotherapy and Cancer Center, National Clinical Research Center for Geriatrics, West China Hospital, Sichuan University, Chengdu 610041, China. Email: xiaweiwei@scu.edu.cn

[#]Authors Lin and Wang contributed equally.

Funding information

National Major Scientific and Technological Special Project, Grant/Award Number: 2018ZX09733001; Development Program of China, Grant/Award Number: 2016YFA0201402; Excellent Youth Foundation of Sichuan Scientific Committee Grant in China, Grant/Award Number: 2019JDJQ008

Abstract

Coronaviruses (CoVs), a subfamily of coronavirinae, are a panel of singlestranded RNA virus. Human coronavirus (HCoV) strains (HCoV-229E, HCoV-OC43, HCoV-HKU1, HCoV-NL63) usually cause mild upper respiratory diseases and are believed to be harmless. However, other HCoVs, associated with severe acute respiratory syndrome, Middle East respiratory syndrome, and COVID-19, have been identified as important pathogens due to their potent infectivity and lethality worldwide. Moreover, currently, no effective antiviral drugs treatments are available so far. In this review, we summarize the biological characters of HCoVs, their association with human diseases, and current therapeutic options for the three severe HCoVs. We also highlight the discussion about novel treatment strategies for HCoVs infections.

KEYWORDS

clinical treatments, COVID-19, human coronaviruses, MERS, SARS

1 | INTRODUCTION

Coronaviruses (CoVs) is a family of enveloped, positive, single-stranded RNA viruses, which are infectious to animals and people, and are able to cause respiratory, hepatic, enteric, and neurological diseases of various severity.^{1,2} Based on their genetic relationship and genomic structures, this family is divided into four genera, termed Alpha-CoV, Beta-CoV, Gamma-CoV, and Delta-CoV.^{3–5} Among the seven identified human coronaviruses, HCoV-NL63 and HCoV-229E belong to the Alpha-CoV, and the other five types are classified as Beta-CoV, including HCoV-OC43, HCoV-HKU1, SARS-CoV, MERS-CoV, and SARS-CoV-2.⁶

It was not until the outbreak of severe acute respiratory syndrome (SARS) in 2002/2003 and Middle East respiratory syndrome (MERS) in 2012 that CoVs were considered as a fatal threat to human beings and received global attention,^{7,8} although they have been discovered for decades.^{9,10} In addition to SARS and MERS, other

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

^{© 2020} The Authors. MedComm published by Sichuan International Medical Exchange & Promotion Association (SCIMEA) and John Wiley & Sons Australia, Ltd.

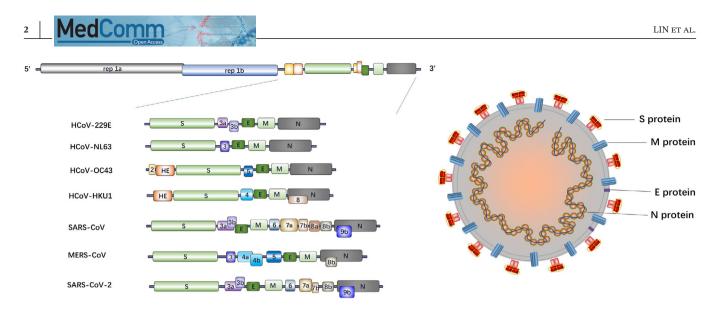


FIGURE 1 Genome organization and structure of HCoVs

human CoVs generally cause only mild upper respiratory diseases, which is similar to common flu.^{11,12} A novel CoV, named SARS-CoV-2 by the World Health Organization (WHO), has emerged again at the end of 2019, causing more infections and deaths worldwide than ever before.¹³ The absence of effective antiviral treatments and serious consequences of these three CoVs have highlighted the urgent need for novel drug development to prevent the spread of CoVs.

Herein, this review mainly focuses on the biological characters of HCoVs, their association with human diseases, and current therapeutic options for the three severe HCoVs. We also highlight the discussion about novel treatment strategies for HCoVs infections.

2 | BIOLOGICAL CHARACTERS OF CORONAVIRUSES

2.1 Genomes

CoVs possess a nonsegmented, positive, single-stranded RNA genome of 26-32 kb.^{2,14,15} All CoVs have a similar genome arrangement with a 5'-methylated cap structure along with 3'-polyadenylated tail. The replicase gene, occupying about 20 kb, two-thirds of the genome and comprising two open reading frames (ORFs), ORF1a and ORF1b, is located at the 5' end.² It encodes two large polyproteins (pp) 1a and 1ab that can be cleaved by papain-like cysteine protease (PLpro) and 3C-like serine protease (3CLpro) into nonstructure proteins, involving some proteases, several RNA modification enzymes, as well as RNA-dependent RNA polymerase (RdRp) and helicase (Hel) required for virus replication.¹⁶ Additionally, an untranslated region (UTR) can also be identified at the 5'-end as same as the 3'terminal. Structure proteins, encompassing the 3'-terminal one-third of the genome, are arranged in a certain order of hemagglutinin esterase (HE) protein that is present in some beta-CoVs, spike protein (S), small membrane protein (E), membrane protein (M), and nucleocapsid protein (N). in brief, the arrangement of the CoV genome can be shown as 5'-UTR-replicase gene (ORF 1a and ORF 1b)-HE protein (if have)-S protein-E protein-M protein-N protein-3' UTR-poly (A)² (Figure 1).

2.2 | Virion structures

CoV is named for the club-shaped projections eradiating from the envelope, which forms its corona or crow-like morphology. The shape of the viral particles is roughly spherical with approximately 80–160 nm in diameters.¹⁷⁻¹⁹ The nucleocapsid protein and the genome RNA intertwine to form a helical structure located inside the envelope. For some CoVs, the spikes on the surface are not only formed by trimers of S protein, but also HE proteins. M protein and E protein, two transmembrane proteins, also participate in the composition of the virus (Figure 1).

S protein, a transmembrane protein, mediates the initiation of CoV infection by attaching to the specific receptors on the target cells.²⁰⁻²² For a prototypical CoV, S protein is usually cleaved into an extracellular receptor binding subunit (S1) and a membrane-anchored subunit (S2), responsible for virus binding and membrane fusion, respectively.^{23,24} Two heptad repeats (HR1 and HR2) and enriched alpha-helices are contained in the S2 domain, a feature typical of fusion protein.²⁵⁻²⁷ The receptor-binding domain (RBD) of S protein specifically binds to target receptors, leading to the conformation change of S1/S2 complex that mediates virus entry.²⁶ Furthermore, the RBD also induces potent neutralizing Ab response, which turns S protein into an important antigenic determinant capable of protective immunity induction and provides a vital approach for the development of immunotherapies.²⁸⁻³²

CoV E protein (8.4-12 kDa) is an integral membrane protein of 76–109 amino acids³³⁻³⁵ and is present in small amount in virions.³⁶⁻³⁸ It contains a short hydrophilic N-terminal followed by a single predicted hydrophobic domain and a hydrophilic C-terminal. Although its membrane topology remains unclear, CoV E protein is commonly referred to as a transmembrane protein with one transmembrane domain.³⁹⁻⁴¹ Accordingly, the E protein mainly targets ER and Golgi-complex and participates in part of the life cycle of the virus, including virus assembly, budding, particles release, envelope formation, and viral pathogenesis.^{33,35,41-50} In addition, CoV E protein may have a crucial role in virus production and maturation, because recombinant CoVs lacking the E protein exhibit significantly reduced viral titers and toxicity.⁵¹⁻⁵⁵

The M protein is a small transmembrane protein (25-30 kDa) with three transmembrane segments, an N-terminal ectodomain and a C terminal-endodomain, determining the shape of the virion.56,57 It is considered as the most abundant structural protein and plays a pivotal role in virion assembly via interacting with other structural proteins.⁵⁸⁻⁶⁰ Binding of S protein and M protein is essential to the virus assembly and the maintenance of S protein in the ER-Golgi intermediate compartment (ERGIC)/Golgi complex.⁶¹⁻⁶³ Virus-like particles (VLPs) are assembled when the combination of M and E proteins occurs, which suggests that they are required for the formation of the envelope.^{39,64-66} Additionally, when expressed alone, it can become a homomultimeric complex, the primary driving force of envelope formation.43,60,67 Furthermore, as to N protein, a stable nucleocapsid and internal core of virions can be achieved when combined with M protein.^{68,69}

The N protein, combined with viral genomic RNA to form a helical nucleocapsid inside the viral envelope, is a multifunctional protein.⁷⁰ It contains three highly conserved domains: the N-terminal domain (NTD), responsible for RNA binding; a C-terminal domain (CTD) that is a hydrophobic and helix-rich terminal, capable of dimerization and oligomerization; and an intrinsically disordered region (RNA-binding domain/domain 2) that is a serine/arginine-rich domain (SR-domain) with a significant phosphorylation potential.^{15,71-86} Phosphorylation of the N protein can initiate structural modification leading to an increased RNA-binding affinity.72,79,87,88 The N protein binds to the genomic RNA through a beads-on-astring form. Likewise, except for the interaction between N protein and nucleic, the ability of complex oligomerization is another pivotal activity required for the formation of the ribonucleoprotein complexes for viral assembly.⁸⁹ In

addition to the role of the N protein in viral core formation and assembly,^{69,90-92} it is also involved in other critical processes of viral life cycle such as virus budding and envelope formation,^{21,93-95} genomic mRNA replication ,and genomic RNA synthesis.^{96,97}

3 | PATHOGENESIS OF CORONAVIRUS IN RESPIRATORY DISEASES

Although the pathogenic mechanisms of human CoVs have not yet been fully understood, the investigation of their unique characteristics of each CoV enables to distinguish the various human CoVs including SARS, MERS, and SARS-COV-2.

3.1 | Target receptors and virus entry

3.1.1 | Target receptors

The crucial early step of the infection of human CoVs into susceptible host cells is the interaction between viral S protein and cellular target receptors. One of the subunits of S protein, S1, containing RBD, is responsible for specific recognition and binding of the target receptors. The other subunit, S2, is in charge of the membrane fusion.^{22,98-102} The tissue tropism, as well as the susceptible host species, is mainly determined by the binding of S protein to target receptors.¹⁰² Based on lines of known evidence, human CoVs utilize multiple and different types of cellular receptors rather than use a common receptor. Therefore, multiple cellular receptors have been identified as target receptors for the various human CoVs to date (Table 1).

Angiotensin-converting enzyme 2 (ACE2), the first known human homolog of Angiotensin-converting enzyme and the receptor for HCoV-NL63, SARS-CoV, and SARS-CoV-2, is a vital component of the reninangiotensin system (RAS).¹⁰³⁻¹⁰⁹ It is a secreted enzyme with a transmembrane domain, a single metalloprotease active site and a signal peptide,¹¹⁰ and predominantly expressed in heart, vascular endothelia, epithelia of the small intestine, kidney, and testis; alveolar macrophage and monocytes of the respiratory tract; and epithelial cells of the trachea, bronchi, and alveoli.^{103,110-112} In contrast to its homolog ACE that contributes to the promotion of lung failure pathogenesis, induction of lung edemas, and impairment of lung function, ACE2 plays a protective role in severe acute lung injury (ALI). Imai et al revealed that the deficiency of ACE2 in the murine models of acute respiratory distress syndrome (ARDS) deteriorated the symptom in lung function, which could be recovered by

dComm Open Acces	and the second	Service of the servic				
	SARS-CoV-2	Beta-CoVs	Bats	Pangolin	ACE2	Heart and vascular endothelia, small-intestine enithelia alvoolar
	MERS-CoV	Beta-CoVs	Bats	Dromedary camel	DPP4	Endothelia cells, epithelial cells, inflammatory cells
	Λ	S		t		l vascular elia, ntestine

coronaviruses
of human
naracteristic
Biological ch
TABLE 1

	HC ₀ V-229E	HCoV-NL63	HCoV-0C43	HC ₀ V-HKU1	SARS-CoV	MERS-CoV	SARS-CoV-2
Genus	Alpha-CoVs	Alpha-CoVs	Beta-CoVs	Beta-CoVs	Beta-CoVs	Beta-CoVs	Beta-CoVs
Natural reservoir	Unknown	Unknown	Unknown	Unknown	Bats	Bats	Bats
Intermediary host	Unknown	Unknown	Unknown	Unknown	Palm civet	Dromedary camel	Pangolin
Target receptor	APN	ACE2	9-O-acetylated sialic acid	9-O-acetylated sialic acid	ACE2	DPP4	ACE2
Receptor distribution	Epithelial cells, endothelial cells, leukocyte, and fibroblast	Heart and vascular endothelia, small intestine epithelia, alveolar macrophage and monocytes, epithelial cells of trachea, bronchi and alveoli	Submaxillary mucin	Submaxillary mucin	Heart and vascular endothelia, small-intestine epithelia, alveolar macrophage and monocytes, epithelial cells of trachea, bronchi and alveoli	Endothelia cells, epithelial cells, inflammatory cells in lung, and smooth muscle cells	Heart and vascular endothelia, small-intestine epithelia, alveolar macrophage and monocytes, epithelial cells of trachea, bronchi and alveoli
Syndromes	Mild upper respiratory diseases, similar to common flu	Mild upper respiratory diseases, similar to common flu	Mild upper respiratory diseases, similar to common flu	Mild upper respiratory diseases, similar to common flu	SARS syndromes	MERS syndromes	COVID-19
Abbreviations: ACE2, an	giotensin-converting enzy	yme 2; APN, aminopeptida	ase N; CoVs, coronaviruse	ss; DPP4, dipeptidyl peptidase	e IV; MERS, Middle East respir	ratory syndrome; SARS, sever	Abbreviations: ACE2, angiotensin-converting enzyme 2; APN, aminopeptidase N; CoVs, coronaviruses; DPP4, dipeptidyl peptidase IV; MERS, Middle East respiratory syndrome; SARS, severe acute respiratory syndrome.

the recombinant ACE2.¹¹³ Thus, downregulation of ACE2 expression in SARS patients could be used as an indicator of severe clinical outcome.¹¹⁴ Besides lung damage caused by SARS-CoV infection could be attenuated by blocking the renin-angiotensin pathway.¹¹⁴ Overall, ACE2 could serve as a novel therapeutic target for severe respiratory diseases.

Dipeptidyl peptidase IV (DPP4), a type II transmembrane protein also known as CD26, is identified as a target receptor for MERS.¹¹⁵ It is a prolyl oligopeptidase expressed in various cells including endothelial cells, epithelial cells, and inflammatory cells in the lung and smooth muscle.¹¹⁶⁻¹¹⁸ The multifunctional protein DDP4 is implicated in the activation of T-cell, the activity regulation of chemokines and growth factors, and the regulation of glucose metabolism.¹¹⁹⁻¹²² Not only can it be embedded in the plasma membrane in the form of a homodimer, but it also presents in extracellular fluid like plasma as a soluble form.^{118,123} Although DDP4 is expressed in epithelial cells of the upper respiratory tract in camels, it is principally found in alveoli but rarely in the nasal cavity or conducting airway.^{115,124} Accordingly, in a murine MERS model, though monocyte infiltration, alveolar edema and microvascular thrombosis were observed in the MERS-CoV-infected lungs, any symptoms were seldom found in the airways.¹²⁵

Aminopeptidase N (APN), a type of II metalloprotease also called CD13, is a ubiquitous enzyme expressed in various organs (lung, intestine, and kidney) and cells (epithelial cells, endothelial cells, leukocyte, and fibroblast).^{126,127} It serves as a target receptor for HCoV-229E,¹²⁸ but not for HCoV-OC43. HCoV-OC43 shares the same specific target with HCoV-HKU1, namely 9-O-acetylated sialic acid.^{129,130}

3.1.2 | Virus attachment and entry

Virus entry is a finely regulated process requiring a series of interactions between the virion and host cell.¹³¹⁻¹³³ Following the conjunction with the target receptor, CoV fuse its envelope with the membrane of the host cell. These processes are forced by the conformational change of S protein, which is triggered by not only the target receptor binding but also PH acidification and proteolytic cleavage led by cell surface or endosomal proteases such as transmembrane protease serine 2 (TMPRSS2), furin, cathepsin L, elastase, and trypsin.¹³⁴⁻¹⁴³ Cleavages of S protein are facilitated at two sites: the boundary between the S1 and S2 subunit (S1/S2) and the conserved site upstream of the fusion peptide (S2').^{138,144} The former one is aimed at releasing RBD from the membrane fusion subunit, and the latter one is important for the exposure of the fusion peptide, hydrophobic in general, which acts as an anchor to target membrane.^{138,145,146} Then the fusion domain adopts two heptad repeats (HR1/HR2) to form a compact coiled-coil conformation called 6-helix bundle or 6HB.^{144,147} Through direct interactions with lipid bilayers, the fusion domain disrupts two apposed membrane layers and fuses the viral envelope to host cell membrane. Ultimately, the viral genome is successfully released into the cytosol of the target cell. In addition to the viral infection through the plasma membrane, the entry of CoVs into cells can be accomplished by the endocytic pathway, depending on the virus strains and host cells.^{135,144}

3.2 | SARS-CoV

SARS-Cov, whose intermediary host is the palm civet, is a highly pathogenic respiratory virus that emerged in 2002, leading to global pandemic that affected more than 8000 people in 29 countries.^{148,149} Patients infected with SARS-CoV initially present "flu-like" syndrome commonly showing high fever, headache, sore throat, myalgia, and dry cough.¹⁵⁰⁻¹⁵² During the disease progression, ALI or ARDS is developed in a number of patients.¹⁵³ Pathological manifestations can be described as three phases. The first step is the disturbance of gas exchange in the first week, owing to the extensive edema, shedding of ciliated epithelial cells, and deposition of hyaline-rich substances on the alveolar membrane. In the next step, pulmonary fibrosis occurs that is characterized as the deposition of fibrin at epithelial cells and the alveolar spaces, as well as the infiltration of inflammatory and fibroblasts. Finally, fibrosis of lung tissue, collagen deposition, and proliferation of alveolar and interstitial cells represent the final step of disease, about 6-8 weeks.¹⁵⁴⁻¹⁵⁸ Diffuse alveolar damage (DAD) accompanied by hyaline membrane formation as well as interstitial thickening is common characteristics of SARS-CoV inducing pulmonary damage.¹⁵⁹ Although many investigators have devoted to inquiring into the pathogenesis of such virus, it has not yet been fully understood until now.

The immune response is the earliest alert system of the host cells warning virus attacks. Ironically, it also aids viral pathogenesis. Pattern recognition receptors (PRRs) that are retinoic acid-inducible gene I protein (RIG-I, member of RLRs family) and melanoma differentiation-associated protein 5 (MDA5) recognize viral pathogen-associated molecular patterns (PAMPs), such as viral components and replication intermediates, to initiate signaling cascades against virus infection.^{160,161} Once the PAMPs from invaded viruses are detected, RIG-I and MDA5 interact with the mitochondrial antiviral signaling protein (MAVs) that is a mitochondrial membrane-bound



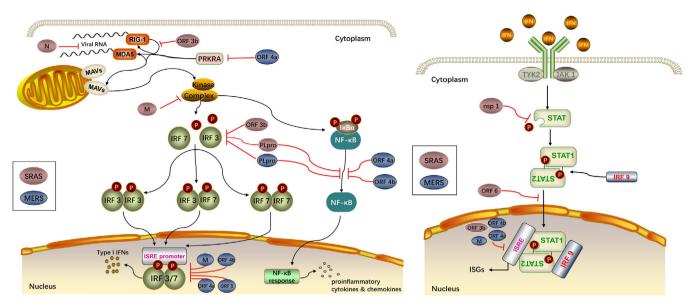


FIGURE 2 Escape mechanisms of innate immune response of SARS-CoV and MERS-CoV

adaptor molecule, followed by the activation of several kinase complexes and multiple subsequent transcription factors (IRF3, IRF7, and NF- κ B). Activation of NF- κ B induces the production of proinflammatory cytokines and chemokines in the nucleus that is a substantial cause of the ARDS.^{162,163} Phosphorylation of IRF3 and IRF7 by the kinase complexes results in homo- or heterodimerization of IRF3 and IRF7. The dimerization initiates the transcription of Type I interferons (IFNs, IFN- α and IFN- β), activating the signal transducer and activator of transcription proteins (STATs) to mediate antiviral response (Figure 2).¹⁶⁴⁻¹⁶⁶

Several defensive approaches are used by SARS-CoV to avoid the PRRs defense system and, ultimately, host innate immune response. One approach is to replicate themselves within the double membrane vesicles (DMVs) that are protected from the PRRs.^{167,168} The eukaryotic mRNAs contain a 5' cap that usually lacks in the viral mRNAs. However, some viruses such as SARS-CoV are capable of building the RNA cap through nsp14 and nsp16/nap10 complex, helping them bypass the recognition by PRRs.¹⁶⁹⁻¹⁷² In addition, a couple of more proteins encoded by the viruses participate in the suppression of the innate immune response by disrupting the IFN response. Among the nonstructural proteins, nsp 1 mainly involves in the degradation of host mRNAs, inactivation of host translational machinery as well as the inhibition of STAT1 phosphorylating.¹⁷³⁻¹⁷⁵ SARS-CoV PLpro interferes phosphorylation of IRF3 and disrupts NF-kB signaling probably via interacting with STING.^{176–178} The nsp7 and nsp15 are also potential IFN antagonists though the mechanism is not clear.¹⁷⁷ Structure proteins such as the N protein and M protein are likely to suppress the type I IFN pathways. Because it was known that N protein inhibits the IFN transcription, the N protein could have a strong potential influence on the viral RNA.¹⁷⁹ M protein blocks the formation of signaling kinase complexes, and suppresses the IRF3 and IRF7 activities, suggesting the potential role of the N protein in the viral infection as well.¹⁸⁰ Such accessory proteins as ORF 3b protein are able to inhibit the RIG-I activity and IRF3 phosphorylation in addition to the transcription of IFN-stimulated genes (ISGs) via the ISRE promoter, while ORF6 blocks the nuclear translocation of STAT1.^{181,182} In spite of the number of research findings in vitro, they have not been validated in vivo. Therefore, it is urgent to examine the findings in vivo for a clear and solid understanding of the infectious process.

Besides the immune response of the host, ACE2 also plays an essential role in the pathogenesis of SARS-CoV. As a negative regulator on the RAS, ACE2 has been closely linked to the pathogenesis of pulmonary diseases and considered as a protective factor for ALI.¹¹³ Consequently, the downregulation of ACE2 mediated by SARS-CoV binding might give an explanation for the progression of severe lung damage occurred on some SARS patients.¹¹⁴

3.3 | MERS-CoV

A decade after SARS, another novel CoV was identified as the pathogen of MERS that caused a higher mortality rate (30%-40%) compared with SARS (around 10%).^{183,184} SARS-CoV and MERS-CoV are both emerged from bats, and are disseminated to human through palm civets and dromedary camels, respectively.¹⁸⁵⁻¹⁸⁸ MERS-CoV and SARS-CoV share some common clinical manifestations ranging from asymptomatic to severe pneumonia in multiple organs,^{150,184,189} and pathological features including inflammatory cells infiltration and DAD.¹⁸³ Similar to SARS-CoV, MERS-CoV is also capable of causing immune dysregulation by attenuating the innate immune response (Figure 2).^{190,191}

Type I interferon is important for the inhibition of MERS-CoV replication in host cells probably via the suppression of the DMVs formation.^{192,193} Capping viral mRNA by nsp14 and nsp16/nap10 complex protects MERS-CoV, as well as SARS-CoV, from the PRRs recognition since the structure of nsp16/nap10 complex in both viruses are analogous.¹⁹⁴ Besides, several proteins of MERS-CoV are involved in immune escape mechanism by involving in the signaling cascades. It was reported that IRF3 nuclear translocation and IFN promoter activation are inhibited by M protein, ORF4a, ORF4b, and ORF5, former three of which also restrained the expression from an the ISRE promoter.¹⁹⁵ Inhibition of the phosphorylation of IRF3 might be the mechanism for IFR3 translocation inhibiting.¹⁹⁴ Moreover, MERS-CoV ORF4a can interact with IFN-inducible double-stranded DNA (dsDNA) dependent protein kinase activator A (PRKRA) and subsequently control the function of RIG-I and MDA5, resulting in the disruption of the IFN response.^{196,197} ORF4a and ORF4b are thought to participate in the NF-*k*B signaling by downregulating the gene stimulated by NF- κ B and affecting the kinase complexes, respectively.¹⁹⁸ Functions of PLpro and nsp1 of MERS-CoV are analogous to the functions of those in SARS-CoV.^{199,200}

Like ACE2, the entry receptor DPP4 for MERS-CoV has also a pivotal role in the disease pathogenesis and is considered as a key factor for the intraspecies variation shown in MERS infection.^{201,202} It is usually expressed in type II pneumocytes that cover 2% of the alveolar surface.^{115,124} Approximately 95% of the surface area is occupied by the type I pneumocytes that are responsible for gas exchange.^{203,204} But the autopsy reports indicated that both type I and II pneumocytes in patients died from MERS-harbored DPP4 expression and these pneumocytes were infected by the virus.^{205,206} MERS-CoV infection of type I pneumocytes might lead to the damage of the cells in the alveoli subsequently causing the DAD.²⁰⁷ It suggests that type I pneumocytes expressing DPP4 might be included in the pathogenesis of the disease. This might explain why chronic obstructive pulmonary disease (COPD) patients attacked by MERS-CoV had poor outcomes, since the expression of DPP4 was predominantly upregulated on type I pneumocytes in such patients.^{202,208} Besides, owing to high expression of DPP4 shown in the kidney, the renal dysfunction might be caused by either the direct infection or the hypoxic damage.¹¹⁶ Evidence of tubular injury, such as cell debris and tubular dilation, could be observed in the late stage of the infection in MERS-CoV-infected mice. However, no virus could be detected in such animals in the early stage after infection, meaning such pathologic changes might be related to the hypoxia.¹²⁵

3.4 | SARS-COV-2

The outbreak of COVID-19, whose pathogen is SARS-CoV-2, now poses a serious threat to the global public health.^{209,210} Since the emergence of the virus, SARS-CoV-2 has affected more than 14 million people with more than 597 thousand deaths worldwide as of July 2020. Next-generation sequencing of the novel virus has been developed soon after the outbreak, indicating that SARS-CoV-2 is closely related to the bat-derived SARSlike CoVs.^{107,211} It is now believed that bats are likely to be the natural reservoir,^{212,213} and pangolins are regarded as intermediary host according to the later studies. Typical clinical presentations of SARS-CoV-2-infected patients include fever, dyspnea, dry cough fatigue, myalgia, pneumonia, and ARDS symptoms, similar to those of SARS and MERS patients.²¹⁴⁻²¹⁶ However, intestinal disorders such as diarrhea are less frequent in COVID-19 patients than SARS.^{214,215,217} Furthermore, in spite of the variation of amino acid at some residues, homology modeling informed that SARS-CoV-2 and SARS-CoV have an analogous RBD structure, and share the same target cell receptor, ACE2, to mediate the viral entry.^{107,108,216,218-220} It is speculated that ACE2 is involved in the pathogenesis of SARS-CoV-2. Owing to the current sparsity of data on the pathological characters of SARS-CoV-2, it is poorly understood. A case report from an infected patient died of this disease showed that DAD with hvaline membrane formation, infiltration of inflammatory cells, and pulmonary edema could be found in the samples taken from their lungs, which is notably corresponding with the symptoms of SARS and MERS patients.²²¹ Additionally, lymphopenia is a common manifestation in COVID-19 patients and might be an effective indicator to estimate the severity of hospitalized COVID-19 patients.²²² Lymphopenia is also supposed to be a vital factor related to the pathogenesis that has not been elucidated so far.²¹³ Moreover, the concentration of some cytokines and chemokines detected in the plasma was higher in COVID-19 patients compared with healthy individuals. Moreover, higher plasma levels of GSCF, IL-2, IL-7, IL-10, MCP1, MIP1A, IP10, and TNF- α were linked to the more severe disease.²¹⁵ All of the data reveal that immunopathology may occupy a crucial place in the development of the disease, and further researches about the pathogenesis of SARS-CoV-2 are urgently needed in the future.

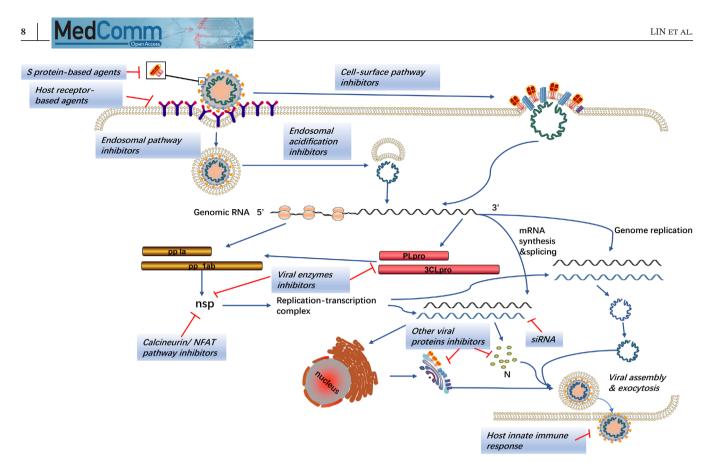


FIGURE 3 Mechanisms of current anti-CoV therapeutic agents

4 | TREATMENTS AND INTERVENTIONS AGAINST CORONAVIRUS

4.1 | Current antivirus therapeutic mechanisms

Owing to the fact that no effective specific antiviral therapies are currently available for SARS, MERS, and COVID-19, isolation and symptomatic support cares are the major management strategies for suspected and confirmed cases requiring hospital treatment including oxygen inhalation, fluid management, and rational use of antibiotics, to prevent organ failure and secondary bacterial infection and alleviate the symptoms.^{189,223-225} Thus, the identification of effective agents against human CoVs is urgently needed in the response to the current COVID-19 outbreak.

All of SARS-CoV, MERS-CoV, and SARS-CoV-2 encode structure proteins (like S protein), nonstructure proteins (eg, PLpro, 3CLpro, RdRp, and helicase), and accessory proteins that are essential for the viral life cycle and that are considered as important targets for the development of antiviral agents (Figure 3).^{226,227} Analyses of genomic sequences and protein structure indicated that the catalytic sites of four crucial enzymes and the key drug-binding pockets in viral enzymes were conserved across SARS-CoV,

MERS-CoV, and SARS-CoV-2.²²⁷ Therefore, the therapeutic experience based on SARS and MERS is capable to guide the treatment of COVID-19.

The idea to disturb the normal life cycle of the virus provides significant insights into the clinical treatment strategy. The S protein is important for the discovery of antiviral agents due to its multifunction in virus infection. RBD located on the S1 subunit can bind to the host cell receptors (ACE2 for SARS-CoV and SARS-CoV-2, DPP4 for MERS-CoV) initiating the conformational changes in S2 subunit to get viral and cell membranes closer and trigger membrane fusion.²²⁸ Consequently, the interaction between RBD and the host cell receptors serves as a key target for the production of neutralizing antibody followed by the vaccine development.^{31,229} Monoclonal antibodies (mAbs) and fusion inhibitors against S1 and S2 subunit, respectively, are potential antivirus drugs to conquer the viral infection, and the agents targeting the host receptors also play a similar role.²³⁰⁻²³⁴ Likewise, cleavage at the protease site of the S1/S2 complex by host proteases such as TMPRSS2 and furin is necessary for the membrane fusion and syncytium formation.^{143,235} The endosomal cysteine protease cathepsins promote the entry of CoVs into the host cell via the endosomal pathway.²³⁶ Inhibitors of these host proteases can potently block the cell entry, which has been proved in vitro and require further validation on animals studies.237

Once entering the host cells, CoVs release the nucleocapsid and genomic RNA into the cytoplasm and start the translation of the replicase gene. The large replicase pp1a and pp1ab are cleaved by PLpro and 3CLpro to produce nonstructural proteins like RdRp and helicase, forming the replication-transcription complex.²³⁸ Numerous agents inhibiting these proteins have shown anti-CoV effects in vitro. Combination of the hydrophobic domains of the replication-transcription complex to the endoplasmic reticulum membrane can form the typical CoV replication structures such as DMVs and convoluted membranes, protecting CoVs from the detection of PRRs.^{168,239} Viral RNA synthesis produces genomic and subgenomic RNAs. Then the subgenomic RNAs are translated to generate the structural and accessory proteins, participating in the assembly of the virion that is released into the extracellular compartment via exocytosis.⁸ Small interfering RNAs (siRNAs) disturbing these processes could be used in the treatment of CoVs infections.

Although CoVs are capable of disturbing the IFN response, they are still sensitive to the IFN treatment in vitro, indicating that augmented host innate IFN response can be an effective strategy to control the viral infection.^{207,240-242} In addition to the enhancement of INF response, several other cell signaling pathways are also regarded as potential anti-CoV targets. These include calcineurin-nuclear factor of activated T cells (NFAT) pathway and kinase signaling pathways such as ERK/MAPK and PI3K/AKT/mTOR pathways, the inhibitors of which have exhibited anti-CoV activities as well.²⁴³⁻²⁴⁵

Since the discovery of new interventions may take months or even years, a more efficient approach is to repurpose existing antiviral agents approved for treating related viral infections. The followings are approved drugs or preclinical compounds that have potential antiviral abilities against SARS, MERS, and COVID-19.

4.2 | Virus-targeted therapeutic strategies

4.2.1 | Agents based on viral enzymes

All of the major proteases of the virus are attractive druggable targets since they are essential for viral transcription and replication (Table 2). As a key part of replication-transcription complex, RdRp participates in the production of genomic RNA and subgenomic RNA. Nucleoside analogues targeting RdRp is capable of inhibiting viral RNA synthesis in a great variety of RNA viruses including CoVs.²⁴⁶⁻²⁵⁰ Favipiravir (T-705), a guanine analogue approved in Japan for influenza treatment, has been proven to effectively interfere the

RNA synthesis of RNA viruses such as influenza virus,

Ebola virus, and other hemorrhagic fever viruses at RdRp level.²⁵¹⁻²⁵⁶ Several studies concluded that favipiravir could inhibit avian influenza A (H5N1) virus and Ebola virus infection in mice.^{256,257} Also, favipiravir has been proved with a notable effect increasing the survival rate of Ebolainfected patients from 35.3% to 56.4% in Sierra Leone.²⁵⁸ A recent study ended with the statement that favipiravir owns the ability against SARS-CoV-2 (EC50 = 61.88uM, CC50 > 400uM, SI > 6.46).²⁵⁹ COVID-19 patients were enrolled in randomized trials for the evaluation of the efficacy of favipiravir plus INF- α or baloxavir marboxil (ChiCTR2000029544 and ChiCTR2000029600). Another guanosine derivative with broad-spectrum antiviral activity, ribavirin, has been authorized for HCV and respiratory syncytial virus (RSV) treatment.²⁶⁰ Accurate mechanism of ribavirin against virus infection is not clear, but inhibition of mRNA capping and viral RNA synthesis could be pivotal to its antiviral activity.²⁶¹ Although high dose of ribavirin has been applied to SARS treatment, the anti-MERS-CoV effects were moderate at such dose in rhesus macaques infected by MERS-CoV and no obvious survival benefit has been observed in MERS patients.^{225,262-267} Recently, an open-label, randomized phase II clinical trial (NCT04276688) has revealed that triple combination of ribavirin, interferon, and lopinavirritonavir in COVID-19 treatment was safe and superior to lopinavir-ritonavir therapy alone in remission of symptoms, shortening virus shedding and promoting discharge of mild to moderate COVID-19 patients.²⁶⁸ Remdesivir (GS-5734) is a small-molecule monophosphoramidate prodrug of an adenosine analog with the ability to interfere with the RNA polymerase and the proofreading exoribonuclease and terminate the nonobligate chain.²⁶⁹⁻²⁷¹ Currently developed for the treatment of Ebola virus infection, remdesivir shows a potential antivirus activity to a diverse panel of RNA viruses including SARS-CoV, MERS-CoV, RSV, Nipah virus (NiV), and Hendra virus in vitro and/or in vivo.270,272,273 Besides, it demonstrated broad-spectrum anti-CoV capacity showing the effective suppression to the epidemic and endemic CoVs, and might be effective against the emerging CoVs now and in the future.²⁷² As to the ongoing COVID-19, clinical and preclinical researches have been set up to investigate the efficacy of remdesivir to the emerging CoV, which indicated that remdesivir was able to inhibit SARS-CoV-2 in cultured cells (EC50 = $0.77 \mu M$ in Vero E6 cells) and a US COVID-19 patient recovered after treating with remdesivir intravenously.^{209,259} Moreover, two phase III clinical trials (NCT04252664 and NCT04257656) planning to enroll 308 and 453 participants, respectively, have been initiated to confirm the value of intravenous remdesivir in COVID-19 patients with the intervention of 200 mg

			~	0								(Continues)
	Refs.	249-257	224,258-265,268	208,257,266-270	257,271	257	248	281	277	278	282-285	(C
	Status	 Randomized trials for COVID-19 in combination with INF-α or baloxavir marboxil (ChiCTR2000029544 and ChiCTR2000029600) 	 Randomized trial for SARS (NCT00578825) Randomized for COVID-19 in combination with a PEGylated interferon (ChiCTR2000029387) Phase II randomized trial for COVID-19 in combination with interferon beta-1b and lopinavir-ritonavir (NCT04276688) 	 Phase III clinical trials for COVID-19 (NCT04252664, NCT04257656, NCT04280705) 	 Phase I clinical trial for Marburg virus (NCT03800173) 	Applied in HSV	Preclinical	Applied in chronic alcohol dependence	Preclinical	Preclinical	 Applied in HIV Phase II/ III clinical trial for MERS (NCT02845843) Phase III clinical trials for COVID-19 (NCT04251871, NCT04255017,NCT0425274, ChiCTR2000029539, ChiCTR2000029308, NCT04261270) 	
	Antiviral spectrum	SARS-CoV-2	SARS-CoV MERS-CoV SARS-CoV-2	SARS-CoV MERS-CoV SARS-CoV-2	SARS-CoV MERS-CoV	SARS-CoV-2	HCoV-NL63 MERS-CoV	SARS-CoV MERS-CoV	SARS-CoV MERS-CoV	MERS-CoV	SARS-CoV MERS-CoV SARS-CoV-2	
	Mechanisms	Interfering with RNA synthesis of RNA virus at RdRp level	Inhibiting mRNA capping and viral RNA synthesis	Interfering the RNA polymerase and the proofreading exoribonuclease and terminating the nonobligate chain	Inhibiting viral RNA polymerase activity via terminating nonobligate RNA chain	RdRp inhibitor	RdRp inhibitor	PLpro inhibitor	Thiopurine analogues that inhibits PLpro	Inhibiting MERS PLpro	3CLpro inhibitors	
Virus-targeted agents for HCoVs	Agents	Favipiravir (T-705)	Ribavirin	Remdesivir (GS-5734)	Galidesivir (BCX4430)	Penciclovir	Acyclovir fleximer analogues	Disulfiram	6-mercaptopurine and 6-thioguanine	Compound 6	Lopinavir / ritonavir	
TABLE 2 Vi	Targets	RdRp						PLpro			3CLpro	

10 MedComm

_
ģ
Ē
÷Ξ
Continue
2
9
7
E 2
LE
BLE
LE

TABLE 2 (Continued)					
Targets	Agents	Mechanisms	Antiviral spectrum	Status	Refs.
	Darunavir and cobicistat	3CLpro inhibitors	SARS-CoV-2	 Applied in HIV Phase III clinical trial for COVID-19 (NCT04252274) 	
	ASC09F	3CLpro inhibitors	SARS-CoV-2	Phase III clinical trial for COVID-19 (NCT04261270)	
	GC376, GC813, GC813, SK80, Peptidomimetic Inhibitors (Compound 6), Neuraminidase inhibitor analogues (compound 3k),	3CLpro inhibitors	SARS-CoV MERS-CoV	• Preclinical	377-381
Helicase	Bananins and 5- Hydroxychromone derivatives	Inhibiting both the ATPase and helicase activities	SARS-CoV	Preclinical	286
	Triazole derivatives (Compound 16)	Inhibiting both the ATPase and helicase activities	MERS-CoV	Preclinical	287
	SSYA10-001 and ADKs	Inhibiting helicase without affecting ATPase activity	SARS-CoV MERS-CoV	Preclinical	288
S protein	Nafamostat	Inhibiting s protein-mediated membrane fusion	MERS-CoV SARS-CoV-2	 Applied in anticoagulant therapy in Asian countries 	257
	Griffithsin	Specially binding to oligosaccharides located on the surface of viral glycoproteins, leading to inhibition of viral entry	HCoV-229E HCoV-OC43 SARS-CoV, MERS-CoV	Preclinical	289,290
	HR2P, HR2P-M2, HR1P, HR1M, HR1L, HR2L, MERS-5HB	Targeting the S2 subunite of S protein of MERS-CoV, thus inhibiting the S protein-mediated membrane fusion	MERS-CoV	Preclinical	227,293,294,382,383
	Peptides (P9)	Inhibiting S protein-mediated membrane fusion	SARS-CoV MERS-CoV	Preclinical	384
	siRNA	Short chains of dsRNA interfering the Narrow-spectrum expression of S protein	Narrow-spectrum	Preclinical	295-297 (Continues)

Targets	Agents	Mechanisms	Antiviral spectrum	Status	Refs.
M, N, E, and accessory proteins	siRNA	Inhibiting the replication of CoVs via Narrow-spectrum silencing M, N, E, ORF3a and ORF7a/7b	Narrow-spectrum	Preclinical	298,299
	Resveratrol	Deregulating the expression of N protein and the apoptosis induced by MERS-CoVs	MERS-CoV	Preclinical	300-306
	Hexamethylene amiloride	Suppressing the ion channel activity of E protein of CoVs	SARS-CoV HCoV-229E	Preclinical	307
DNA metabolism	Gemcitabine hydrochloride	Inhibitor of DNA metabolism	SARS-CoV MERS-CoV	Preclinical	308
Lipid membrane	LJ001 and JL103	Inhibiting membrane fusion via mediating the changes of membrane properties, including the decrease in membrane fluidity	Broad-spectrum against enveloped viruses	Preclinical	309
Host cell membrane-bound viral replication complex	K22	Inhibiting membrane-bound RNA synthesis and double membrane vesicle formation	HCoV-229E SARS-CoV MERS-CoV	Preclinical	385
Others	Arbidol	Unclear	SARS-CoV-2	 Phase IV clinical trials for COVID-19 (NCT04260594, NCT04254874, NCT04255017) 	I
	Oseltamivir	Unclear	SARS-CoV-2	 Phase IV clinical trial for COVID-19 (NCT04255017) Phase III clinical trial for COVID-19 (NCT04261270) 	1

Abbreviations: CoV, coronavirus; MERS, Middle East respiratory syndrome; PLpro, papain-like cysteine protease; RdRp, RNA-dependent RNA polymerase; SARS, severe acute respiratory syndrome; 3CLpro, 3C-like serine protease.

12

on day 1 and 100 mg once-daily maintenance for 9 days. However, the first clinical trial (NCT04252664) has been suspended so far and the second trial (NCT04257656) with 237 COVID-19 patients enrolled finally indicated that remdesivir hardly shown any statistically significant clinical benefits.²⁷⁴ Conversely, a research found that 36 of 53 (68%) hospitalized patients suffered from severe COVID-19 and treated with compassionate-use remdesivir could achieve clinical improvement.²⁷⁵ In addition, a phase III, randomized, double-blind, placebo-controlled trial (NCT04280705) conducted by Beigel et al uncovered the fact that remdesivir prevailed over placebo in shortening the time to recovery in adults patients.²⁷⁶ Though remdesivir has been approved by the Food and Drug Administration (FDA) to treat severe COVID-19 patients, further researches are urgently required to determine the efficacy and the indication of remdesivir therapy. A novel synthesized nucleoside analogue, BCX4430 (Galidesivir), is designed to inhibit viral RNA polymerase activity via terminating nonobligate RNA chain.²⁷⁷ BCX4430 exhibits a promising antiviral capability against a wide array of RNA viruses including filoviruses (Ebola virus and Marburg virus) and CoVs (SARS-CoV and MERS-CoV).²⁷⁷ It is currently tested in phase I clinical trial (NCT03800173) for Marburg virus and can be a potential countermeasure against viral diseases that threaten the public health in the world. A recent study also concluded that penciclovir, another RdRp inhibitor that is approved for HSV, showed effects on reducing SARS-CoV-2 infection by high-dose administration (EC50 = 95.96 μ M, CC50 > 400 μ M, SI > 4.17).²⁵⁹ Although resistance to nucleoside analogs has rarely been reported, it is worth noting that mutation in RdRp is probably responsible for the acquired resistance and should be monitored for the possible resistance.²⁷⁸

CoVs PLpro enzymes display proteolytic, deubiquitylating, and deISGylating activities.²⁷⁹⁻²⁸¹ PLpro was first regarded as a druggable target for SARS-CoV, and then several compounds targeting SARS-CoV PLpro were also found to be effective against MERS-CoV PLpro, recently.^{282,283} Though numerous PLpro inhibitors have been identified, many of them only inhibit enzymatic activities and do not affect on the viral replication.^{284,285} A study from Lin et al suggested that an FDA-approved alcoholaversive drug, disulfiram could inhibit SARS-CoV and MERS-CoV PLpro via different mechanisms. And the synergistic inhibition between disulfiram and known PLpro inhibitors, like 6-thioguanine and mycophenolic acid, to MERS-CoV might offer the potential combination treatments against CoVs in clinical.²⁸⁶

Another essential protease that cleaves the viral polyproteins during viral replication is 3CLpro. Similar to PLpro, a great many of inhibitors have been identified with the ability to target CoVs 3CLpro. Among the 3CLpro **MedComm**

inhibitors, the human immunodeficiency virus (HIV) protease inhibitors are the most comprehensively studied such as lopinavir, ritonavir, ASC09F, as well as darunavir and cobicistat. Lopinavir and ritonavir, applied in combination to treat HIV infection, have shown improvement in the outcome of SARS patients in nonrandomized trials.^{287,288} Though lopinavir alone hardly displayed antiviral activity against MERS-CoV in vitro, the combination of lopinavir and ritonavir ameliorated the outcome in MERS-CoV-infected nonhuman primates.^{289,290} Therefore, the efficacy of the lopinavir-ritonavir combination in MERS patients should be reappraised (NCT02845843). However, no benefit was observed in lopinavir-ritonavir treatment compared to standard care in a randomized, controlled, open-label clinical trial (ChiCTR2000029308) involving severe COVID-19 patients.²⁹¹ Further trials are still needed to confirm the therapeutic efficacy. In addition, several other clinical trials have been developed to confirm the efficacy of 3CLpro inhibitors on COVID-19 (NCT04252274, NCT04251871, NCT04255017, ChiCTR2000029539, NCT04251871, NCT04255017, and NCT04261270), as well as darunavir and cobicistat (NCT04252274), ASC09F combined with oseltamivir (NCT04261270).

Helicase acts on the duplex oligonucleotides and turns them into single strands in an ATP-dependent manner in the CoV replication cycle. That helicases in different CoVs are highly homologous making them potentially strong targets for the CoVs therapeutic options. Based on the mechanism actions, the helicase inhibitors can be approximately classified into two groups. One is able to inhibit both the ATPase and helicase activities represented by bananins derivatives, 5-hydroxychromone derivatives, and triazole derivatives (compound 16).^{292,293} The other group including SSYA 10-001 and ADKs has the ability to inhibit the helicase activity with no or little effects on the ATPase activity.²⁹⁴ However, the toxicity of helicase inhibitors needs to be examined before being applied to human patients.

4.2.2 | Agents based on viral structure proteins

The transmembrane glycoprotein, S protein, is also a promising target for antiviral agents' development (Table 2). One class of antiviral drugs targeting S protein mostly blocks the spike-mediated membrane fusion. A potent MERS-CoV inhibitor, nafamostat, has demonstrated to be inhibitive against the SARS-CoV-2 infection (EC50 = 22.50 μ M, CC50 > 100 μ M, SI > 4.44).²⁵⁹ Griffithsin is a red-alga-derived lectin, which specially binds to oligosaccharides located on the surface of viral glycoproteins, for example, S glycoprotein of SARS-CoV and HIV glycoprotein 120.295 A wide range of human CoVs infection could be inhibited by griffithsin, comprising HCoV-229E, HCoV-NL63, HCoV-OC43, and SARS-CoV.^{295,296} But the value of griffithsin in the treatment or prevention of COVID-19 is needed to be evaluated urgently. S2 subunit of S protein contains two heptad repeat (HR1 and HR2). Antiviral peptides analogous derived from these regions exhibited inhibition to the spike protein-mediated cell-cell fusion and viral entry in viruses such as SARS-CoV, MERS-CoV, as well as HCoV-229E.^{231,297,298} HR2P, a peptide spanning HR2 sequences of MERS-CoV S protein, was capable of interacting with HR1 region to form a 6-HB complex, resulting in potent inhibition of viral fusion and replication. Its analog HR2P-M2 exhibited an obvious improvement in stability, solubility, and antiviral activity after being modified with hydrophilic residues.²²⁸ Additionally, HR2P-M2 intranasal administration effectively prevented experimental mice transduced by adenoviral vectors conveying human DPP4 from MERS-CoV infection with a >1000-fold decrease in viral titers in the lung, and this protection was intensified via the combination of HR2P-M2 and IFN- β .²⁹⁹ Another newly designed fusion inhibitor from MERS-CoV called MERS-five-helix bundle (MERS-5HB), which is derived from the 6-HB, also displayed a potent suppression on S protein-mediated syncytial formation. Compared with MERS-6HB, MERS-5HB lacks one HR2, which endows its capability to interact with viral HR2 to interrupt the membrane fusion.³⁰⁰ Besides, MERS-5HB could effectively inhibit the entry of pseudotyped MERS-CoV with 50% inhibitory concentration (IC50) about 1 µM.³⁰⁰ Altogether, the resistance of these drugs can be overcome by combining antiviral peptides aiming at various domains of S2 subunit, which may also attain synergistic effects in vitro. As for siRNAs, which displayed antiviral activities in vitro as well as in SARS-CoV-infected rhesus macaques, are still under preclinical development and demand further studies to seek out the reliable drug delivery methods in a human before the clinical application.³⁰¹⁻³⁰³

M, N, and E proteins and some accessory proteins are not only vital to the virion assembly but also involved in viral pathogenesis in which they function in the interruption of host innate immune response to assist viral infection. For instance, M protein as well as accessory proteins 4a, 4b, and 5 of MERS-CoV act as IFN antagonist, and N protein of SARS-CoV serves as an inhibitor of viral RNA. Researches carried out by He et al and Akerstrom et al emphasized that siRNAs silencing M, N, E, ORF3a, and ORF7a/7b play an important role in the inhibition of viral replication of the SARS-CoV.^{304,305} But the delivery methods still limit their application in human being. Nevertheless, various agents related to these proteins are discovered via functional analysis. One example is resveratrol, a natural stilbene derivative demonstrated to reduce inflammation and exert antiviral activity against multiple viruses.³⁰⁶⁻³¹¹ In addition, it exhibited significant inhibition of MERS-CoV infection and prolonged cellular survival after virus challenge in vitro via deregulating the expression of N protein and the apoptosis induced by MERS-CoV.³¹² Alternatively, hexamethylene amiloride, a viroporin inhibitor, is capable to suppress the ion channel activity of E protein of CoVs such as HCoV-229E and SARS-CoV.³¹³ Identified as DNA metabolism inhibitor, gemcitabine hydrochloride is a deoxycytidine analog inhibiting both SARS-CoV and MERS-CoV with micromolar EC50 and low toxicity, which suggests its potential antiviral capacity for other human CoVs.³¹⁴ LJ001 and JL103, two novel lipophilic thiazolidine derivatives, could induce several changes in membrane properties including the decrease in membrane fluidity, contributing to inhibition of membrane fusion, which made them become broad-spectrum enveloped virus entry inhibitors and potential anti-CoV agents.315

4.3 | Host-cell-targeted therapies

4.3.1 | Enhancement of host innate immune response

The host innate immune response is vital to the interruption of viral replication. Recombinant interferons have been applicated in treating various viruses as well as many CoVs (Table 3). Though the host interferon response can be inhibited by the CoVs, they are still proved effective against CoVs infection such as SARS-CoV and MERS-CoV in vitro and several animal models.^{242,264,289,299,316} Recombinant interferons are usually combined with other antiviral agents including ribavirin or lopinavir/ritonavir to treat SARS-CoV and MERS-CoV infections, 290,317 and the anti-CoVs efficacy of interferons is enhanced when added with ribavirin.³¹⁸ A combination of interferon- α 2b with ribavirin reduced virus replication and improves clinical outcomes in a rhesus macaque model of MERS.²⁶⁴ However, the effectiveness of combination treatments comprising interferons and ribavirin is still controversial in clinical researches. A study of five patients received interferon- $\alpha 2b$ and ribavirin showed no survival, but the finding might be not reliable owing to the delayed administration.²⁶⁶ Contrarily, in another study (n = 44), mortality rates of individuals receiving interferon- α 2b and ribavirin exhibited a significant reduction in day 14, compared with the individuals received standard supportive care, but no significant difference was observed in day 28.265 Moreover, no significant difference in mortality rates between interferon- α 2b and ribavirin treatment group and interferon- β 1a and ribavirin

TABLE 3 Host-targeted agents for HCoVs



Targets	Agents	Mechanisms	Antiviral spectrum	Status	Refs.
Host inter- feron response	Recombinant interferons	Exogenous interferons augmented host innate IFN response against CoVs infection	SARS-CoV MERS-CoV SARS-CoV-2	 Marked Randomized trial for COVID-19 (NCT04251871, ChiCTR2000029638) 	284,285,311,:
	Nitazoxanide	Inducing the host innate immune response via intensifying interferon- α and interferon- β production through fibroblasts and activation of protein kinase R (PKR)	SARS-CoV-2	• Marked	313-315
	Polyinosinic:polycytidylic acid (poly(I:C))	Type I interferon inducer	MERS-CoV	 Phase II clinical trial for malignant gliomas patients Preclinical research for MERS-CoV 	191,316,317
Host receptor	N-(2-aminoethyl)-1- aziridine-ethanamine (NAAE), P4 and P5	ACE2 inhibitors blocking cell-cell membrane fusion and viral entry	SARS-CoV SARS-CoV-2	• Marked	318,319
	YS110 and Anti-DPP4 mAb clones 2F9	DPP4 inhibitors blocking cell-cell membrane fusion and viral entry	MERS-CoV	 Phase I clinical trial of YS110 for patients with advance solid tumors Preclinical research of mAbs clones 2F9 for MERS-CoV 	320
Host proteases	K11777	Cathepsin inhibitors inhibiting the endosomal pathway, thus blocking viral entry	SARS-CoV MERS-CoV	Preclinical	236,321,322
	Camostat mesylate	TMPRSS2 inhibitor interrupting the TMPRSS2-mediated cell surface entry	HCoV-229E SARS-CoV MERS-CoV	Preclinical	323,324
	dec-RVKR-CMK	Furin inhibitor inhibiting furin-mediated cleavage of S	MERS-CoV	Preclinical	143
Endocytosis	Chlorpromazine, triflupromazine, fluphenazine	Inhibiting clathrin-mediated endocytosis via affecting the assembly of clathrin-coated pits at the plasma membrane	SARS-CoV MERS-CoV	• Approved as antipsychotic agents	308,326
	Ouabain and bufalin	Binding sodium/potassium- transporting ATPase subunit α1 (ATP1A1)	MERS-CoV	• Approved as cardiotonic steroids	326

(Continues)

Targets	Agents	Mechanisms	Antiviral spectrum	Status	Refs.
	Chloroquine, Hydroxychloroquine	Increasing endosomal pH, thus affecting viral infection	HCoV-229E HCoV-OC43 SARS-CoV MERS-CoV SARS-CoV-2	 Approved as autoimmune disease and antimalarial agents Open-label trial for COVID-19 (ChiCTR2000029609) Phase II clinical trial of chloroquine for COVID-19 (NCT04323527) Phase III clinical trial of hydroxychloroquine for COVID-19 (NCT04334148) 	257,327-333
Other antiviral agents	Cyclosporin A (CsA), alisporivir	Cyclophilin inhibitors affecting the calcineurin/NFAT pathway, blocking the viral replication	HCoV-NL63 HCoV-229E SARS-CoV MERS-CoV	• Marked	243,244,335-33
	Selumetinib, Trametinib, Rapamycin	Inhibiting the ERK/MAPK and PI3K/AKT/mTOR signaling pathways	SARS-CoV MERS-CoV	• Marked	242
	Dexamethasone, Methylprednisolone	-	SARS-CoV MERS-CoV SARS-CoV-2	 Marked Clinical trials for COVID-19 patients (ChiCTR2000029386, ChiCTR2000029656, ChiCTR2000029656, NCT0424459) 	350-355

Abbreviations: ACE2, Angiotensin-converting enzyme 2; CoV, coronavirus; DPP4, dipeptidyl peptidase IV; SARS, severe acute respiratory syndrome; MERS, Middle East respiratory syndrome.

treating group was observed in a retrospective study.²⁶⁷ On the other hand, interferon- β 1b displayed stronger inhibition to the MERS-CoV replication in vitro compared with other interferons, and combined use of interferon- β 1b with other antiviral compounds should be evaluated in further research.^{289,290}

Nitazoxanide, originally identified as an antiprotozoal agent, was later considered as a broad-spectrum antiviral agent able to inhibit the replication of numerous DNA and RNA viruses such as RSV, parainfluenza, rotavirus, HBV, HCV, HIV, yellow fever, as well as CoVs in vitro.³¹⁹ Several clinical trials have confirmed its potential value in treating influenza, chronic HBV, and HCV.³¹⁹⁻³²¹ Moreover, recent research indicated that nitazoxanide was capable of inhibiting SARS-CoV-2 infection at a low micromolar concentration (EC50 = 2.12 μ M; CC50 > 35.53 μ M; SI > 16.76), and the in vivo evaluation of this efficacy is demanded.²⁵⁹ Another type I interferon inducer,

Polyinosinic:polycytidylic acid (poly(I:C)), is an analog of dsDNA with a powerful ability to reduce MERS-CoV load in animal models.¹⁹² And phase II clinical trials demonstrated that it was well tolerated by malignant gliomas patients when injected intramuscularly.^{322,323} Overall, the use of interferons or interferon inducers may be a valuable strategy against CoVs infection and should be furtherly accessed in animal models.

4.3.2 | Antiviral agents based on host entry factors

Several host factors are considered essential to CoVs entry, such as the host receptors that bind to CoVs S protein, and host proteases that facilitated CoVs entry through the cell surface or endosomal pathway. Thus, these factors become attractive targets for anti-CoV agents' development (Table 3). Antibodies, peptides, and some functional inhibitors targeting the host receptors can effectively interrupt the binding between S protein and the host cells. One example is that the N-(2-aminoethyl)-1-aziridine-ethanamine (NAAE), a small molecular inhibitor, is able to target ACE2 leading to the block of S protein-mediated membrane fusion, so does the synthetic peptides analogous, P4 and P5.^{324,325} Similar agents were also found in MERS treatment, one of which, YS110, was confirmed to be well tolerated in patients with advance solid tumors.³²⁶ Owing to their specificity to appointed receptors, they were regarded as narrow-spectrum drugs. However, the efficacy of these receptor-targeted agents have never been evaluated in any CoVs-infected patients and the safeties of these agents also remain unclear.

Host protease such as cathepsins and TMPRSS2 play a key role in the cleavage of S protein and the suppression of these proteases with potent inhibitors can obstruct the virus entry through either endosomal pathway or cell surface pathway. K11777 is a cathepsin inhibitor with broad spectrum against enveloped RNA virus including SARS-CoV and MERS-CoV.237,327,328 However, the camostat mesylate, applied in chronic pancreatitis treating, is a TMPRSS2 inhibitor that interrupts the TMPRSS2mediated cell surface entry.^{329,330} The combination of cathepsin inhibitors and TMPRSS2 inhibitors is crucial to the complete suppression of MERS-CoV in vitro. Inhibition of another host protease, furin, which is vital to furinmediated S cleavage for CoVs, also can block membrane fusion and the viral entry like MERS-CoV.¹⁴³ Notably, inhibition of host proteases may result in some side effects and need further evaluation.

Some approved antipsychotic agents (chlorpromazine, triflupromazine, and fluphenazine) and cardiotonic steroids (ouabain and bufalin) can inhibit clathrinmediated endocytosis via affecting the assembly of clathrin-coated pits at the plasma membrane and binding sodium/potassium-transporting ATPase subunit α 1 (ATP1A1), respectively.^{314,331,332} Thus, they are considered as clathrin-mediated endocytosis inhibitors. Alternatively, endosomal acidification also has a profound impact on endocytosis. Increase of endosomal pH shows an inhibitive effect on virus infection, which has been confirmed by chloroquine, a widely used autoimmune disease and antimalarial agents.333 Chloroquine proves to be active against a wide range of RNA viruses including HCoV-229E, HCoV-OC43, SARS-CoV, and MERS-CoV.³³⁴⁻³³⁸ Recently, chloroquine is identified to function at both entry and postentry phase of the SARS-CoV-2 infection with the EC90 value of 6.90 µM in Vero E6 cells.²⁵⁹ Additionally, as an immunoregulator, its antiviral activity may be synergistically intensified in vivo.²⁵⁹ Therefore, chloroquine is suggested as a potential

MedComm

option against the emerging SARS-CoV-2. Significantly, higher dose of chloroquine should not be recommended for severe COVID-19 patients owing to the its drug safety, particularly while simultaneously accepting azithromycin and oseltamivir treatment, which was presented by a randomized clinical trial (NCT04323527).339 Hydroxychloroquine, a chloroquine analog with lower toxicity, was listed as a potential anti-SARS-CoV-2 agent and recommended to be applied in COVID-19 treatment by Chinese national guideline and FDA.340 However, evidence of the benefits and harms of hydroxychloroquine therapy is still insufficient and conflicting. Some small studies show that hydroxychloroquine was capable of decreasing SARS-CoV-2 shedding that could be enhanced by the combination of azithromycin and achieving a shorter time to clinical recovery.^{341,342} But almost no clinical benefit was observed in other studies.^{340,343} Therefore, therapeutic efficacy of hydroxychloroquine is still needed to be reconfirmed. Moreover, there are several factors required to be reconsidered before efficacy evaluation, such as the severity of illness, definition of the endpoints, and effects of the comorbidities.

4.3.3 | Other antiviral agents

Except for the innate immune response, host receptors, and other factors affecting the endocytosis, some signaling pathways have also been suggested as useful approaches for discovering anti-CoV reagents (Table 3). Cyclophilins are peptidyl-prolyl isomerases with physiological functions showing as modulating the calcineurin/NFAT pathway via reacting with CoVs nsp1, which is important for the immune cell activation.²⁴⁴ In addition, they are also required for the replication of numerous viruses including HIV, HCV, as well as CoVs.³⁴⁴ Consequently, inhibition of cyclophilins by cyclosporines, such as cyclosporin A (CsA) and its derivatives, has shown to block the replication of a wide range of CoVs.244,245,345,346 However, the obvious immune suppressive properties of CsA limit its application in antiviral therapy. But alisporivir, a nonimmunosuppressive cyclosporin A-analog, also displayed the inhibition to the CoVs replication at a lowmicromolar concentration.³⁴⁷ Additionally, the combined use of cyclosporine and interferon or ribavirin in vitro was beneficial to inhibit SARS-CoV or MERS-CoV infection, which needed to be furtherly evaluated in animal models.^{347,348} Furthermore, some antiviral agents inhibiting the cellular signaling pathways, in particular, the ERK pathway and PI3K/AKT pathway, also interrupt the replication of CoVs.^{243,314,349} However, the efficacy and safety against CoVs infection of these agents still need to be reconsidered.

Corticosteroids, which were used in SARS and MERS treatment, have been linked to several complications such as psychosis, diabetes, and avascular necrosis.^{350,351} They also were pointed out to be associated with viral replication prolongation in MERS patients.³⁵¹ However, an update on the efficacy of dexamethasone based on a press release publicized recently indicated that severe COVID-19 patients given 6 mg dexamethasone once daily shown a lower mortality (about 8-26%) compared to patients with standard care.^{352,353} Besides, another agent, methylprednisolone, also exhibited potential capacity in reducing the mortality rate and achieving better clinical outcomes in severe COVID-19 patients.^{354,355} Thus, it is wise to apply corticosteroids to the right patients at the right time. But physicians also need to monitor the corticosteroid-related complications. Clinical trials of corticosteroid treatments are shown in Table 3.

4.4 | Potential immunotherapeutic options

4.4.1 | Antibody and plasma therapy

S protein of SARS-CoV proves to be highly immunogenic during infection and responsible for eliciting a humoral immune response in the host.³⁵⁶ Antivirus antibodies could be detected in the plasma of convalescent patients' infected SARS-CoV and MERS-CoV.357,358 Convalescent plasma therapy has been applicated in treating patients infected by numerous viruses involving Ebola virus, Junin virus, Machupo virus, and Lassa fever.³⁵⁹⁻³⁶² As for SARS-CoV, higher day-22 discharge rate and lower mortality rate have been observed among SARS patients who received convalescent plasma transfusion before day 14 of the illness.^{363,364} This is consistent with another small cohorts research concluding infected patients with severe conditions who failed to respond to current therapies but finally survived after transfused with convalescent plasma, with no obvious side effects.³⁵⁸ Similar results were found in MERS patients.³⁶⁵ Additionally, plasma adoptive therapy with anti-MERS-CoV antibodies could block the virus adhesion and accelerate the viral elimination from MERS-CoV-infected animal models.¹⁹² But the efficacy and safety of convalescent plasma therapy in COVID-19 patients still need to be reevaluated. Although convalescent plasma therapy proves to be a potentially effective therapeutic option for emerging CoVs, several factors still limit its extensive use in clinical, one of which is enough titers of serum neutralizing antibodies.

The development of mAbs targeting the S protein of CoVs is regarded as a remedial strategy (Table 4). Potent mAbs against S protein of human CoVs can be

attained via transgenic mice immunization, convalescent B cells immortalization, and cloning of small chain variable regions from naïve and convalescent patients.³⁶⁶ The majority of mAbs interact with the RBD of S protein leading to the interruption of RBD-receptor binding and block the viral attachment. A few mAbs react with other regions of S protein besides the RBD.³⁶⁶ Although binding to different epitopes, these mAbs exhibit capacity to reduce the viral titers. Two RBD-specific neutralizing mAbs, MERS-4 and MERS-27, which were derived from single-chain variable regions, revealed suppressive effects against both MERS-CoV and pseudotyped MERS-CoV infection at nanomolar concentrations and were recommended as promising candidates for therapeutic interventions to MERS.²³² Based on similar mechanisms, other human mAbs for MERS-CoV were also capable of competing with DPP4 for RBD binding and neutralizing the virus.^{233,234,367-370} When administrated to individuals at risk, some of the mAbs were capable of preventing viral replication and contributing to block the transmission of MERS-CoV among human.³⁶⁸ Thus, such antibodies could be served as prophylactic strategies in clinical and valuable tools to guild the development of effective anti-CoVs vaccines.234,368

4.4.2 | Vaccine

From SARS-CoV to SARS-CoV-2, the emergence of severe human CoVs have taught us many lessons about the importance of rapid diagnostics and effective vaccines to control the outbreak caused by these viruses. Due to the persistence of zoonotic sources in endemic areas, lethal CoVs remain existing in human society and may lead to the epidemic at any time. Thus, a priority is to develop vaccines targeting conserved alleles and providing broad-spectrum protection against varied viral strains. Since the emergence of SARS-CoV and MERS-CoV, several strategies were applicated in vaccine design, including inactivated virus vaccines, live-attenuated virus vaccines, viral vector vaccines, nanoparticles, recombinant protein subunits vaccines, and DNA vaccines (Table 4).371,372 And clinical trials have also been developed to test the efficacy of the novel vaccines (Table 5). Effective vaccines are pivotal in blocking the virus spread from animals' reservoirs to human hosts. Inactivated virus vaccines, preserving the viral structure and antigenicity but eliminating the infectious ability, could elicit neutralizing antibodies in animal models of SARS-CoV and show protection against viral replication when administrated with or without adjuvants.372-375 Different from inactivated virus vaccines, live attenuated virus vaccines are generated via reducing the virulence of live viruses, meaning that they are still able to induce infection,



TABLE 4	Potential	immunothera	peutic o	ptions for HCoVS

Therapeutic	ential minutotherapeute options for freevs	Antiviral		
options	Design	spectrum	Status	Refs.
Convalescent plasma	Plasma from convalescent patients infected with viruses	Broad-spectrum SARS-CoV MERS-CoV SARS-CoV-2	Clinical trials	346,351-353
MERS-4, MERS-27, m336, m337, m338, REGN3051 and REGN3048	Targeting S protein and attaining via transgenic mice immunization, convalescent B cells immortalization, and cloning of small chain variable regions from naïve and convalescent patients	SARS-CoV MERS-CoV	 Preclinical Phase I clinical trial for MERS 	231-233,361-364
Inactivated virus vaccines	Rendering the viral genome noninfectious with chemicals or radiation while maintaining the virion structure	SARS-CoV MERS-CoV SARS-CoV-2	 Preclinical Phase I/ II clinical trial for COVID-19 	365,366
live- attenuated virus vaccines	Reducing or eliminating the virulence of a live virus with chemical-driven or site-directed mutagenesis	SARS-CoV MERS-CoV	• Preclinical	365,366
Viral vector vaccines	Viral vectors (MVA or Ad) that expressing full-length or the S1 subunit of S protein	SARS-CoV MERS-CoV SARS-CoV-2	 Preclinical Phase I/ II clinical trial for COVID-19 and MERS 	365,366
Nanoparticles	Purified S protein-containing nanoparticles produced in insect cells, which were infected with specific recombinant baculovirus containing the gene encoding S protein of CoVs	SARS-CoV MERS-CoV SARS-CoV-2	 Preclinical Phase I clinical trial for COVID-19 	386
Recombinant protein subunits vaccines	Antigenic components either full-length S protein or RBD subunit of S protein of CoVs	SARS-CoV MERS-CoV SARS-CoV-2	 Preclinical Phase I clinical trial for COVID-19 	365,366
DNA vaccines	DNA encoding viral antigenic components	SARS-CoV MERS-CoV SARS-CoV-2	 Preclinical Phase I clinical trial for SARS Phase I/ II clinical trial for COVID-19 and MERS 	365,366

Abbreviations: CoV, coronavirus; MERS, Middle East respiratory syndrome; SARS, severe acute respiratory syndrome.

which may be related to the disseminated infection observed in immunocompromised patients. In addition, live-attenuated virus vaccines can induce an innate and adaptive immune response and the protective value can last for a long time.³⁷¹ Besides, other strategies for vaccines development are also evaluated in animal models. Based on the experience of SARS-CoV and MERS-CoV, the development of novel SARS-CoV-2 vaccine is currently underway and requires more research. However, several concerns should be addressed about the vaccination. The first is the disease deterioration caused by vaccination. Although this situation only appears in a small subset of SARS vaccine studies, it is still a significant problem that needs to be properly solved.³⁷² Second, the variability of S protein can mediate CoVs escape from neutralization, suggesting that recombinant protein subunits vaccines based on S protein may demand multivalent approaches.³⁷⁶ Last but not least, how to define

19

TABLE 5	Important clinical	trials with vac	cines for SARS	S-CoV, MERS-C	CoV, and SARS-CoV-2	
HCoVs	Trial	Phase	Status	Sample size	Types	Design
SARS-CoV	NCT00099463	Phase I	Completed	10	DNA vaccine	A recombinant DNA vaccine, VRC-SRSDNA015-00-VP.
SARS-CoV-2	NCT04299724	Phase I	Recruiting	100	Viral vector vaccine	Pathogen-specific aAPC
	NCT04276896	Phase I/ II	Recruiting	100	Viral vector vaccine	LV-SMENP DC and antigen-specific cytotoxic T cell vaccines
	NCT04283461	Phase I	Recruiting	45	Nanoparticles mRNA-based vaccine	mRNA-1273
	NCT04352608	Phase I/ II	Recruiting	744	Inactivated vaccine	SARS-CoV-2-inactivated vaccine
	NCT04412538	Phase I/ II	Recruiting	942	Inactivated vaccine	SARS-CoV-2-inactivated vaccine
	NCT04405908	Phase I	Recruiting	150	Recombinant S protein subunits vaccines	A recombinant SARS-CoV-2 trimeric S protein subunit vaccine, SCB 2019
	NCT04445389	Phase I/ II	Recruiting	190	DNA vaccine	A DNA vaccine, GX-19
	NCT04368728	Phase I/ II	Recruiting	7600	RNA vaccine	RNA vaccine, BNT162a1, BNT162b1, BNT162b2, BNT162c1
	NCT04368988	Phase I	Recruiting	131	Nanoparticle vaccine	A SARS-CoV-2 recombinant spike protein nanoparticle vaccine (SARS-CoV-2 rS) with Or without MATRIX-M [™] adjuvant
	NCT04453852	Phase I	Recruiting	40	Recombinant protein subunits vaccines	Recombinant protein SARS-COV-2 vaccine, Covax-19
MERS-CoV	NCT03399578	Phase I	Recruiting	48	Viral vector vaccine	ChAdOx1 MERS
	NCT03615911	Phase I	Completed	26	Viral vector vaccine	MVA-MERS-S
	NCT04170829	Phase I	Recruiting	24	Viral vector vaccine	ChAdOx1 MERS
	NCT04130594	Phase I/ II	Recruiting	162	Viral vector vaccine	BVRS-GamVac
	NCT04128059	Phase I/ II	Recruiting	268	Viral vector vaccine	BVRS-GamVac-Combi
	NCT04119440	Phase I	Not yet recruiting	160	Viral vector vaccine	MVA-MERS-S_DF1
	NCT02788188	Phase I	Completed	38	mAbs	SAB-301
	NCT03721718	Phase I/ II	Active, not recruiting	60	DNA vaccine	GLS-5300
	NCT02670187	Phase I	Completed	75	DNA vaccine	GLS-5300

the target individuals suitable for the vaccination. It is recommended that people at a high risk of CoV exposure such as health care workers should be vaccinated.³⁷⁷

5 | CONCLUSION AND OUTLOOK

MedComm

The emergence and prevalence of highly pathogenic CoV severely threaten public health. A task of top priority is to make clear the viral structural and epidemiological characteristics and block the viral dissemination as well as the progression of the disease, at the first case. To date, further understanding of the life cycle and the pathogenesis of emerging human CoVs makes current therapeutic strategies of antiviral infection more rational. Repurposing existing antiviral drugs is an effective short-term strategy to deal with emerging CoV such as the ongoing SARS-CoV-2. Various agents with different targets have been evaluated in vitro and in vivo. But not all antiviral agents are capable of achieving better efficacy than in vitro, and in vivo studies are needed to select optimal agents. Suitable animal models are particularly significant. However, there are only a few effective animal models available for the studies of CoVs treatment, which may postpone the clinical evaluation of drugs. Besides, due to the diversity of viruses and the capacity of rapidly mutating, some antiviral reagents available for existing CoVs may become invalid. Accordingly, sequencing more natural specimens and combination therapy with two or more synergistic agents have become promising solutions. Furthermore, the efficacy of current antiviral therapies needs to be estimated in larger scale, well-organized randomized clinical trial

The efficacy and safety of the most practicable strategies for the ongoing COVID-19 are urgently needed to be further assessed in clinical trials, involving remdesivir (GS-5734), lopinavir/ritonavir, lopinavir/ritonavir combined with IFN- β , convalescent plasma, and mAbs, which prove to be potentially effective options against SARS-CoV-2. Additionally, to control the future CoV pandemics, the development of novel broad-spectrum antiviral agents covering numerous CoVs will become the ultimate goal.

ACKNOWLEDGMENTS

This work is supported by the National Major Scientific and Technological Special Project for "Significant New Drugs Development" (No. 2018ZX09733001, China), the Development Program of China (No. 2016YFA0201402), and by the Excellent Youth Foundation of Sichuan Scientific Committee Grant in China (No. 2019JDJQ008).

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ORCID

Xiawei Wei D https://orcid.org/0000-0002-6513-6422

REFERENCES

- 1. Woo PCY, Lau SKP, Huang Y, Yuen K-Y. Coronavirus diversity, phylogeny and interspecies jumping. *Exp Biol Med (Maywood)*. 2009;234:1117-1127.
- Weiss SR, Leibowitz JL. Coronavirus pathogenesis. Adv Virus Res. 2011;81:85-164.
- Chan JFW, Li KSM, To KKW, Cheng VCC, Chen H, Yuen K-Y. Is the discovery of the novel human betacoronavirus 2c EMC/2012 (HCoV-EMC) the beginning of another SARS-like pandemic? *J Infect*. 2012;65:477-489.
- 4. Woo PCY, Lau SKP, Lam CSF, et al. Discovery of seven novel mammalian and avian coronaviruses in the genus deltacoronavirus supports bat coronaviruses as the gene source of alphacoronavirus and betacoronavirus and avian coronaviruses as the gene source of gammacoronavirus and deltacoronavirus. *J Virol.* 2012;86:3995-4008.
- 5. Chan JF-W, To KK-W, Tse H, Jin D-Y, Yuen K-Y. Interspecies transmission and emergence of novel viruses: lessons from bats and birds. *Trends Microbiol.* 2013;21:544-555.
- Jimenez-Guardeño JM, Nieto-Torres JL, DeDiego ML, et al. The PDZ-binding motif of severe acute respiratory syndrome coronavirus envelope protein is a determinant of viral pathogenesis. *PLoS Pathog.* 2014;10:e1004320-e.
- 7. Zhong NS, Zheng BJ, Li YM, et al. Epidemiology and cause of severe acute respiratory syndrome (SARS) in Guang-

dong, People's Republic of China, in February, 2003. *Lancet*. 2003;362:1353-1358.

- 8. Chan JFW, Lau SKP, To KKW, Cheng VCC, Woo PCY, Yuen K-Y. Middle East respiratory syndrome coronavirus: another zoonotic betacoronavirus causing SARS-like disease. *Clin Microbiol Rev.* 2015;28:465-522.
- 9. Tyrrell DA, Bynoe ML. Cultivation of viruses from a high proportion of patients with colds. *Lancet*. 1966;1:76-77.
- Hamre D, Procknow JJ. A new virus isolated from the human respiratory tract. *Proc Soc Exp Biol Med*. 1966;121:190-193.
- Su S, Wong G, Shi W, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trends Microbiol*. 2016;24:490-502.
- Forni D, Cagliani R, Clerici M, Sironi M. Molecular evolution of human coronavirus genomes. *Trends Microbiol*. 2017;25:35 -48.
- Ryu S, Chun BC. Korean Society of Epidemiology -nCo VTFT. An interim review of the epidemiological characteristics of 2019 novel coronavirus. *Epidemiol Health*. 2020;42:e2020006-e.
- Liu DX, Yuan Q, Liao Y. Coronavirus envelope protein: a small membrane protein with multiple functions. *Cell Mol Life Sci.* 2007;64:2043-2048.
- Lai MM, Cavanagh D. The molecular biology of coronaviruses. Adv Virus Res. 1997;48:1-100.
- Fang SG, Shen H, Wang J, Tay FPL, Liu DX. Proteolytic processing of polyproteins 1a and 1ab between non-structural proteins 10 and 11/12 of coronavirus infectious bronchitis virus is dispensable for viral replication in cultured cells. *Virology*. 2008;379:175-180.
- Neuman BW, Adair BD, Yoshioka C, et al. Supramolecular architecture of severe acute respiratory syndrome coronavirus revealed by electron cryomicroscopy. *J Virol.* 2006;80:7918-7928.
- Bárcena M, Oostergetel GT, Bartelink W, et al. Cryo-electron tomography of mouse hepatitis virus: insights into the structure of the coronavirion. *Proc Natl Acad Sci USA*. 2009;106:582-587.
- Saif LJ. Coronavirus immunogens. Vet Microbiol. 1993;37:285-297.
- Song HC, Seo M-Y, Stadler K, et al. Synthesis and characterization of a native, oligomeric form of recombinant severe acute respiratory syndrome coronavirus spike glycoprotein. *J Virol.* 2004;78:10328-10335.
- Siu YL, Teoh KT, Lo J, et al. The M, E, and N structural proteins of the severe acute respiratory syndrome coronavirus are required for efficient assembly, trafficking, and release of viruslike particles. *J Virol.* 2008;82:11318-11330.
- 22. Kirchdoerfer RN, Cottrell CA, Wang N, et al. Pre-fusion structure of a human coronavirus spike protein. *Nature*. 2016;531:118-121.
- 23. Holmes KV. SARS coronavirus: a new challenge for prevention and therapy. *J Clin Invest*. 2003;111:1605-1609.
- 24. Hofmann H, Pöhlmann S. Cellular entry of the SARS coronavirus. *Trends Microbiol*. 2004;12:466-472.
- Ingallinella P, Bianchi E, Finotto M, et al. Structural characterization of the fusion-active complex of severe acute respiratory syndrome (SARS) coronavirus. *Proc Natl Acad Sci U S A*. 2004;101:8709-8714.
- 26. Liu S, Xiao G, Chen Y, et al. Interaction between heptad repeat 1 and 2 regions in spike protein of SARS-associated coronavirus:

implications for virus fusogenic mechanism and identification of fusion inhibitors. *Lancet*. 2004;363:938-947.

- Tripet B, Howard MW, Jobling M, Holmes RK, Holmes KV, Hodges RS. Structural characterization of the SARS-coronavirus spike S fusion protein core. *J Biol Chem.* 2004;279:20836-20849.
- Godet M, Grosclaude J, Delmas B, Laude H. Major receptorbinding and neutralization determinants are located within the same domain of the transmissible gastroenteritis virus (coronavirus) spike protein. *J Virol.* 1994;68:8008-8016.
- Kubo H, Yamada YK, Taguchi F. Localization of neutralizing epitopes and the receptor-binding site within the aminoterminal 330 amino acids of the murine coronavirus spike protein. *J Virol.* 1994;68:5403-5410.
- Bonavia A, Zelus BD, Wentworth DE, Talbot PJ, Holmes KV. Identification of a receptor-binding domain of the spike glycoprotein of human coronavirus HCoV-229E. J Virol. 2003;77:2530-2538.
- He Y, Zhou Y, Liu S, et al. Receptor-binding domain of SARS-CoV spike protein induces highly potent neutralizing antibodies: implication for developing subunit vaccine. *Biochem Biophys Res Commun.* 2004;324:773-781.
- He Y, Lu H, Siddiqui P, Zhou Y, Jiang S. Receptor-binding domain of severe acute respiratory syndrome coronavirus spike protein contains multiple conformation-dependent epitopes that induce highly potent neutralizing antibodies. *J Immunol.* 2005;174:4908-4915.
- 33. Raamsman MJ, Locker JK, de Hooge A, et al. Characterization of the coronavirus mouse hepatitis virus strain A59 small membrane protein E. *J Virol*. 2000;74:2333-2342.
- 34. Arbely E, Khattari Z, Brotons G, Akkawi M, Salditt T, Arkin IT. A highly unusual palindromic transmembrane helical hairpin formed by SARS coronavirus E protein. *J Mol Biol.* 2004;341:769-779.
- Kuo L, Hurst KR, Masters PS. Exceptional flexibility in the sequence requirements for coronavirus small envelope protein function. J Virol. 2007;81:2249-2262.
- Liu DX, Inglis SC. Association of the infectious bronchitis virus 3c protein with the virion envelope. *Virology*. 1991;185:911-917.
- Tung FY, Abraham S, Sethna M, et al. The 9-kDa hydrophobic protein encoded at the 3' end of the porcine transmissible gastroenteritis coronavirus genome is membrane-associated. *Virology*. 1992;186:676-683.
- Yu X, Bi W, Weiss SR, Leibowitz JL. Mouse hepatitis virus gene 5b protein is a new virion envelope protein. *Virology*. 1994;202:1018-1023.
- Corse E, Machamer CE. Infectious bronchitis virus E protein is targeted to the Golgi complex and directs release of virus-like particles. J Virol. 2000;74:4319-4326.
- Sharpe HJ, Stevens TJ, Munro S. A comprehensive comparison of transmembrane domains reveals organelle-specific properties. *Cell*. 2010;142:158-169.
- Nieto-Torres JL, Dediego ML, Alvarez E, et al. Subcellular location and topology of severe acute respiratory syndrome coronavirus envelope protein. *Virology*. 2011;415:69-82.
- An S, Chen CJ, Yu X, Leibowitz JL, Makino S. Induction of apoptosis in murine coronavirus-infected cultured cells and demonstration of E protein as an apoptosis inducer. *J Virol.* 1999;73:7853-7859.

- de Haan CA, Vennema H, Rottier PJ. Assembly of the coronavirus envelope: homotypic interactions between the M proteins. *J Virol.* 2000;74:4967-4978.
- Wilson L, McKinlay C, Gage P, Ewart G. SARS coronavirus E protein forms cation-selective ion channels. *Virology*. 2004;330:322-331.
- Nal B, Chan C, Kien F, et al. Differential maturation and subcellular localization of severe acute respiratory syndrome coronavirus surface proteins S, M and E. J Gen Virol. 2005;86:1423-1434.
- Yang Y, Xiong Z, Zhang S, et al. Bcl-xL inhibits T-cell apoptosis induced by expression of SARS coronavirus E protein in the absence of growth factors. *Biochem J*. 2005;392:135-143.
- 47. Machamer CE, Youn S. The transmembrane domain of the infectious bronchitis virus E protein is required for efficient virus release. *Adv Exp Med Biol.* 2006;581:193-198.
- Yuan Q, Liao Y, Torres J, Tam JP, Liu DX. Biochemical evidence for the presence of mixed membrane topologies of the severe acute respiratory syndrome coronavirus envelope protein expressed in mammalian cells. *FEBS Lett.* 2006;580:3192-3200.
- Ye Y, Hogue BG. Role of the coronavirus E viroporin protein transmembrane domain in virus assembly. J Virol. 2007;81:3597-3607.
- DeDiego ML, Nieto-Torres JL, Jiménez-Guardeño JM, et al. Severe acute respiratory syndrome coronavirus envelope protein regulates cell stress response and apoptosis. *PLoS Pathog.* 2011;7:e1002315-e.
- Curtis KM, Yount B, Baric RS. Heterologous gene expression from transmissible gastroenteritis virus replicon particles. J Virol. 2002;76:1422-1434.
- Ortego J, Escors D, Laude H, Enjuanes L. Generation of a replication-competent, propagation-deficient virus vector based on the transmissible gastroenteritis coronavirus genome. *J Virol.* 2002;76:11518-11529.
- Kuo L, Masters PS. The small envelope protein E is not essential for murine coronavirus replication. *J Virol.* 2003;77:4597-4608.
- DeDiego ML, Alvarez E, Almazán F, et al. A severe acute respiratory syndrome coronavirus that lacks the E gene is attenuated in vitro and in vivo. *J Virol.* 2007;81:1701-1713.
- 55. Ortego J, Ceriani JE, Patiño C, Plana J, Enjuanes L. Absence of E protein arrests transmissible gastroenteritis coronavirus maturation in the secretory pathway. *Virology*. 2007;368:296-308.
- Voss D, Kern A, Traggiai E, et al. Characterization of severe acute respiratory syndrome coronavirus membrane protein. *FEBS Lett.* 2006;580:968-973.
- Armstrong J, Niemann H, Smeekens S, Rottier P, Warren G. Sequence and topology of a model intracellular membrane protein, El glycoprotein, from a coronavirus. *Nature*. 1984;308:751-752.
- Kuo L, Masters PS. Genetic evidence for a structural interaction between the carboxy termini of the membrane and nucleocapsid proteins of mouse hepatitis virus. J Virol. 2002;76:4987-4999.
- Masters PS. The molecular biology of coronaviruses. *Adv Virus Res.* 2006;66:193-292.
- Neuman BW, Kiss G, Kunding AH, et al. A structural analysis of M protein in coronavirus assembly and morphology. *J Struct Biol.* 2011;174:11-22.

- Opstelten DJ, Raamsman MJ, Wolfs K, Horzinek MC, Rottier PJ. Envelope glycoprotein interactions in coronavirus assembly. *J Cell Biol.* 1995;131:339-349.
- Mortola E, Roy P. Efficient assembly and release of SARS coronavirus-like particles by a heterologous expression system. *FEBS Lett.* 2004;576:174-178.
- 63. Fehr AR, Perlman S. Coronaviruses: an overview of their replication and pathogenesis. *Methods Mol Biol.* 2015;1282:1-23.
- Vennema H, Godeke GJ, Rossen JW, et al. Nucleocapsidindependent assembly of coronavirus-like particles by co-expression of viral envelope protein genes. *EMBO J*. 1996;15:2020-2028.
- Baudoux P, Carrat C, Besnardeau L, Charley B, Laude H. Coronavirus pseudoparticles formed with recombinant M and E proteins induce alpha interferon synthesis by leukocytes. J Virol. 1998;72:8636-8643.
- 66. Corse E, Machamer CE. The cytoplasmic tails of infectious bronchitis virus E and M proteins mediate their interaction. *Virology*. 2003;312:25-34.
- 67. Lim KP, Liu DX. The missing link in coronavirus assembly. Retention of the avian coronavirus infectious bronchitis virus envelope protein in the pre-Golgi compartments and physical interaction between the envelope and membrane proteins. J Biol Chem. 2001;276:17515-17523.
- 68. Narayanan K, Maeda A, Maeda J, Makino S. Characterization of the coronavirus M protein and nucleocapsid interaction in infected cells. *J Virol.* 2000;74:8127-8134.
- Escors D, Ortego J, Laude H, Enjuanes L. The membrane M protein carboxy terminus binds to transmissible gastroenteritis coronavirus core and contributes to core stability. *J Virol.* 2001;75:1312-1324.
- Sturman LS, Holmes KV, Behnke J. Isolation of coronavirus envelope glycoproteins and interaction with the viral nucleocapsid. *J Virol.* 1980;33:449-462.
- Parker MM, Masters PS. Sequence comparison of the N genes of five strains of the coronavirus mouse hepatitis virus suggests a three domain structure for the nucleocapsid protein. *Virology*. 1990;179:463-468.
- Masters PS. Localization of an RNA-binding domain in the nucleocapsid protein of the coronavirus mouse hepatitis virus. *Arch Virol.* 1992;125:141-160.
- Huang Q, Yu L, Petros AM, et al. Structure of the N-terminal RNA-binding domain of the SARS CoV nucleocapsid protein. *Biochemistry*. 2004;43:6059-6063.
- Surjit M, Liu B, Kumar P, Chow VTK, Lal SK. The nucleocapsid protein of the SARS coronavirus is capable of selfassociation through a C-terminal 209 amino acid interaction domain. *Biochem Biophys Res Commun.* 2004;317:1030-1036.
- Chang C-K, Sue S-C, Yu T-H, et al. The dimer interface of the SARS coronavirus nucleocapsid protein adapts a porcine respiratory and reproductive syndrome virus-like structure. *FEBS Lett.* 2005;579:5663-5668.
- Fan H, Ooi A, Tan YW, et al. The nucleocapsid protein of coronavirus infectious bronchitis virus: crystal structure of its N-terminal domain and multimerization properties. *Structure*. 2005;13:1859-1868.
- 77. Surjit M, Kumar R, Mishra RN, Reddy MK, Chow VTK, Lal SK. The severe acute respiratory syndrome coronavirus nucleocapsid protein is phosphorylated and localizes in the cyto-

plasm by 14-3-3-mediated translocation. *J Virol*. 2005;79:11476-11486.

- Yu IM, Gustafson CLT, Diao J, et al. Recombinant severe acute respiratory syndrome (SARS) coronavirus nucleocapsid protein forms a dimer through its C-terminal domain. *J Biol Chem*. 2005;280:23280-23286.
- Chang C-K, Sue S-C, Yu T-H, et al. Modular organization of SARS coronavirus nucleocapsid protein. J Biomed Sci. 2006;13:59-72.
- Spencer K-A, Hiscox JA. Characterisation of the RNA binding properties of the coronavirus infectious bronchitis virus nucleocapsid protein amino-terminal region. *FEBS Lett.* 2006;580:5993-5998.
- Chen C-Y, Chang C-K, Chang Y-W, et al. Structure of the SARS coronavirus nucleocapsid protein RNA-binding dimerization domain suggests a mechanism for helical packaging of viral RNA. J Mol Biol. 2007;368:1075-1086.
- Peng T-Y, Lee K-R, Tarn W-Y. Phosphorylation of the arginine/serine dipeptide-rich motif of the severe acute respiratory syndrome coronavirus nucleocapsid protein modulates its multimerization, translation inhibitory activity and cellular localization. *FEBS J.* 2008;275:4152-4163.
- Hurst KR, Koetzner CA, Masters PS. Identification of in vivointeracting domains of the murine coronavirus nucleocapsid protein. *J Virol.* 2009;83:7221-7234.
- Wu C-H, Yeh S-H, Tsay Y-G, et al. Glycogen synthase kinase-3 regulates the phosphorylation of severe acute respiratory syndrome coronavirus nucleocapsid protein and viral replication. *J Biol Chem.* 2009;284:5229-5239.
- Lo Y-S, Lin S-Y, Wang S-M, et al. Oligomerization of the carboxyl terminal domain of the human coronavirus 229E nucleocapsid protein. *FEBS Lett.* 2013;587:120-127.
- Takeda M, Chang C-K, Ikeya T, et al. Solution structure of the c-terminal dimerization domain of SARS coronavirus nucleocapsid protein solved by the SAIL-NMR method. *J Mol Biol.* 2008;380:608-622.
- Nelson GW, Stohlman SA, Tahara SM. High affinity interaction between nucleocapsid protein and leader/intergenic sequence of mouse hepatitis virus RNA. *J Gen Virol.* 2000;81:181-188.
- Luo H, Chen Q, Chen J, Chen K, Shen X, Jiang H. The nucleocapsid protein of SARS coronavirus has a high binding affinity to the human cellular heterogeneous nuclear ribonucleoprotein A1. *FEBS Lett.* 2005;579:2623-2628.
- Zlotnick A. Theoretical aspects of virus capsid assembly. *J Mol Recognit.* 2005;18:479-490.
- He R, Dobie F, Ballantine M, et al. Analysis of multimerization of the SARS coronavirus nucleocapsid protein. *Biochem Biophys Res Commun.* 2004;316:476-483.
- Chen H, Gill A, Dove BK, et al. Mass spectroscopic characterization of the coronavirus infectious bronchitis virus nucleoprotein and elucidation of the role of phosphorylation in RNA binding by using surface plasmon resonance. *J Virol.* 2005;79:1164-1179.
- 92. de Haan CA, Smeets M, Vernooij F, Vennema H, Rottier PJ. Mapping of the coronavirus membrane protein domains involved in interaction with the spike protein. *J Virol.* 1999;73:7441-7452.
- Boscarino JA, Logan HL, Lacny JJ, Gallagher TM. Envelope protein palmitoylations are crucial for murine coronavirus assembly. *J Virol.* 2008;82:2989-2999.

24 | MedComm

- Ruch TR, Machamer CE. The hydrophobic domain of infectious bronchitis virus E protein alters the host secretory pathway and is important for release of infectious virus. *J Virol*. 2011;85:675-685.
- 95. Ruch TR, Machamer CE. The coronavirus E protein: assembly and beyond. *Viruses*. 2012;4:363-382.
- Sims AC, Ostermann J, Denison MR. Mouse hepatitis virus replicase proteins associate with two distinct populations of intracellular membranes. *J Virol.* 2000;74:5647-5654.
- Stertz S, Reichelt M, Spiegel M, et al. The intracellular sites of early replication and budding of SARS-coronavirus. *Virology*. 2007;361:304-315.
- Walls AC, Tortorici MA, Bosch B-J, et al. Cryo-electron microscopy structure of a coronavirus spike glycoprotein trimer. *Nature*. 2016;531:114-117.
- Walls AC, Tortorici MA, Frenz B, et al. Glycan shield and epitope masking of a coronavirus spike protein observed by cryoelectron microscopy. *Nat Struct Mol Biol.* 2016;23:899-905.
- Gui M, Song W, Zhou H, et al. Cryo-electron microscopy structures of the SARS-CoV spike glycoprotein reveal a prerequisite conformational state for receptor binding. *Cell Res.* 2017;27:119-129.
- 101. Pallesen J, Wang N, Corbett KS, et al. Immunogenicity and structures of a rationally designed prefusion MERS-CoV spike antigen. *Proc Natl Acad Sci U S A*. 2017;114:E7348-E57.
- 102. Yuan Y, Cao D, Zhang Y, et al. Cryo-EM structures of MERS-CoV and SARS-CoV spike glycoproteins reveal the dynamic receptor binding domains. *Nat Commun.* 2017;8:15092.
- 103. Tipnis SR, Hooper NM, Hyde R, Karran E, Christie G, Turner AJ. A human homolog of angiotensin-converting enzyme. Cloning and functional expression as a captopril-insensitive carboxypeptidase. *J Biol Chem.* 2000;275:33238-33243.
- Li W, Moore MJ, Vasilieva N, et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature*. 2003;426:450-454.
- 105. Hofmann H, Pyrc K, van der Hoek L, Geier M, Berkhout B, Pöhlmann S. Human coronavirus NL63 employs the severe acute respiratory syndrome coronavirus receptor for cellular entry. *Proc Natl Acad Sci U S A*. 2005;102:7988-7993.
- 106. Hofmann H, Simmons G, Rennekamp AJ, et al. Highly conserved regions within the spike proteins of human coronaviruses 229E and NL63 determine recognition of their respective cellular receptors. *J Virol.* 2006;80:8639-8652.
- 107. Lu R, Zhao X, Li J, et al. Genomic characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding. *Lancet*. 2020;395(10224):565-574.
- 108. Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor recognition by novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS. *J Virol.* 2020;94(7):e00127-20.
- 109. Imai Y, Kuba K, Ohto-Nakanishi T, Penninger JM. Angiotensin-converting enzyme 2 (ACE2) in disease pathogenesis. *Circ J*. 2010;74:405-410.
- Donoghue M, Hsieh F, Baronas E, et al. A novel angiotensinconverting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1–9. *Circ Res.* 2000;87:E1-E9.
- 111. Harmer D, Gilbert M, Borman R, Clark KL. Quantitative mRNA expression profiling of ACE 2, a novel homologue of angiotensin converting enzyme. *FEBS Lett.* 2002;532:107-110.

- 112. Hamming I, Timens W, Bulthuis MLC, Lely AT, Navis GJ, van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus. A first step in understanding SARS pathogenesis. *J Pathol.* 2004;203:631-637.
- Imai Y, Kuba K, Rao S, et al. Angiotensin-converting enzyme 2 protects from severe acute lung failure. *Nature*. 2005;436:112-116.
- 114. Kuba K, Imai Y, Rao S, et al. A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury. *Nat Med.* 2005;11:875-879.
- 115. Meyerholz DK, Lambertz AM. Dipeptidyl peptidase 4 distribution in the human respiratory tract: implications for the middle east respiratory syndrome. *Am J Pathol.* 2016;186:78-86.
- 116. Lambeir A-M, Durinx C, Scharpé S, De Meester I. Dipeptidylpeptidase IV from bench to bedside: an update on structural properties, functions, and clinical aspects of the enzyme DPP IV. *Crit Rev Clin Lab Sci.* 2003;40:209-294.
- 117. Deacon CF. Physiology and pharmacology of DPP-4 in glucose homeostasis and the treatment of type 2 diabetes. Front Endocrinol (Lausanne). 2019;10:80.
- 118. Xu J, Wang J, He M, et al. Dipeptidyl peptidase IV (DPP-4) inhibition alleviates pulmonary arterial remodeling in experimental pulmonary hypertension. *Lab Invest.* 2018;98:1333-1346.
- 119. Reinhold D, Bank U, Bühling F, et al. Inhibitors of dipeptidyl peptidase IV (DP IV, CD26) induces secretion of transforming growth factor-beta 1 (TGF-beta 1) in stimulated mouse splenocytes and thymocytes. *Immunol Lett.* 1997;58:29-35.
- Barnett A. DPP-4 inhibitors and their potential role in the management of type 2 diabetes. *Int J Clin Pract.* 2006;60:1454-1470.
- 121. Ohnuma K, Dang NH, Morimoto C. Revisiting an old acquaintance: cD26 and its molecular mechanisms in T cell function. *Trends Immunol.* 2008;29:295-301.
- 122. Mortier A, Gouwy M, Van Damme J, Proost P, Struyf S. CD26/dipeptidylpeptidase IV-chemokine interactions: doubleedged regulation of inflammation and tumor biology. *J Leukoc Biol.* 2016;99:955-969.
- Klemann C, Wagner L, Stephan M, von Hörsten S. Cut to the chase: a review of CD26/dipeptidyl peptidase-4's (DPP4) entanglement in the immune system. *Clin Exp Immunol.* 2016;185:1-21.
- 124. Widagdo W, Raj VS, Schipper D, et al. Differential Expression of the middle east respiratory syndrome coronavirus receptor in the upper respiratory tracts of humans and dromedary camels. *J Virol.* 2016;90:4838-4842.
- 125. Li K, Wohlford-Lenane C, Perlman S, et al. Middle east respiratory syndrome coronavirus causes multiple organ damage and lethal disease in mice transgenic for human dipeptidyl peptidase 4. *J Infect Dis.* 2016;213:712-722.
- 126. Luan Y, Xu W. The structure and main functions of aminopeptidase N. *Curr Med Chem.* 2007;14:639-647.
- 127. Yeager CL, Ashmun RA, Williams RK, et al. Human aminopeptidase N is a receptor for human coronavirus 229E. *Nature*. 1992;357:420-422.
- Wentworth DE, Holmes KV. Molecular determinants of species specificity in the coronavirus receptor aminopeptidase N (CD13): influence of N-linked glycosylation. *J Virol.* 2001;75:9741-9752.

MedComn

- 129. Huang X, Dong W, Milewska A, et al. Human Coronavirus HKU1 spike protein uses O-acetylated sialic acid as an attachment receptor determinant and employs hemagglutininesterase protein as a receptor-destroying enzyme. *J Virol.* 2015;89:7202-7213.
- 130. Hulswit RJG, Lang Y, Bakkers MJG, et al. Human coronaviruses OC43 and HKU1 bind to 9-O-acetylated sialic acids via a conserved receptor-binding site in spike protein domain A. *Proc Natl Acad Sci U S A*. 2019;116:2681-2690.
- 131. Marsh M, Helenius A. Virus entry: open sesame. *Cell*. 2006;124:729-740.
- 132. Grove J, Marsh M. The cell biology of receptor-mediated virus entry. *J Cell Biol*. 2011;195:1071-1082.
- Boulant S, Stanifer M, Lozach P-Y. Dynamics of virus-receptor interactions in virus binding, signaling, and endocytosis. *Viruses*. 2015;7:2794-2815.
- 134. Simmons G, Reeves JD, Rennekamp AJ, Amberg SM, Piefer AJ, Bates P. Characterization of severe acute respiratory syndrome-associated coronavirus (SARS-CoV) spike glycoprotein-mediated viral entry. *Proc Natl Acad Sci U S A*. 2004;101:4240-4245.
- 135. Matsuyama S, Ujike M, Morikawa S, Tashiro M, Taguchi F. Protease-mediated enhancement of severe acute respiratory syndrome coronavirus infection. *Proc Natl Acad Sci U S A*. 2005;102:12543-12547.
- 136. Simmons G, Gosalia DN, Rennekamp AJ, Reeves JD, Diamond SL, Bates P. Inhibitors of cathepsin L prevent severe acute respiratory syndrome coronavirus entry. *Proc Natl Acad Sci U S A*. 2005;102:11876-11881.
- 137. Qiu Z, Hingley ST, Simmons G, et al. Endosomal proteolysis by cathepsins is necessary for murine coronavirus mouse hepatitis virus type 2 spike-mediated entry. *J Virol.* 2006;80:5768-5776.
- Belouzard S, Chu VC, Whittaker GR. Activation of the SARS coronavirus spike protein via sequential proteolytic cleavage at two distinct sites. *Proc Natl Acad Sci U S A*. 2009;106:5871-5876.
- Belouzard S, Madu I, Whittaker GR. Elastase-mediated activation of the severe acute respiratory syndrome coronavirus spike protein at discrete sites within the S2 domain. *J Biol Chem.* 2010;285:22758-22763.
- 140. Bertram S, Glowacka I, Müller MA, et al. Cleavage and activation of the severe acute respiratory syndrome coronavirus spike protein by human airway trypsin-like protease. *J Virol.* 2011;85:13363-13372.
- 141. Glowacka I, Bertram S, Müller MA, et al. Evidence that TMPRSS2 activates the severe acute respiratory syndrome coronavirus spike protein for membrane fusion and reduces viral control by the humoral immune response. *J Virol*. 2011;85:4122-4134.
- 142. Shulla A, Heald-Sargent T, Subramanya G, Zhao J, Perlman S, Gallagher T. A transmembrane serine protease is linked to the severe acute respiratory syndrome coronavirus receptor and activates virus entry. *J Virol.* 2011;85:873-882.
- 143. Millet JK, Whittaker GR. Host cell entry of Middle East respiratory syndrome coronavirus after two-step, furin-mediated activation of the spike protein. *Proc Natl Acad Sci U S A*. 2014;111:15214-15219.

- Belouzard S, Millet JK, Licitra BN, Whittaker GR. Mechanisms of coronavirus cell entry mediated by the viral spike protein. *Viruses*. 2012;4:1011-1033.
- 145. Heald-Sargent T, Gallagher T. Ready, set, fuse! The coronavirus spike protein and acquisition of fusion competence. *Viruses*. 2012;4:557-580.
- 146. Millet JK, Whittaker GR. Physiological and molecular triggers for SARS-CoV membrane fusion and entry into host cells. *Virology*. 2018;517:3-8.
- 147. White JM, Delos SE, Brecher M, Schornberg K. Structures and mechanisms of viral membrane fusion proteins: multiple variations on a common theme. *Crit Rev Biochem Mol Biol.* 2008;43:189-219.
- 148. Song Z, Xu Y, Bao L, et al. From SARS to MERS, thrusting coronaviruses into the spotlight. *Viruses*. 2019;11:59.
- 149. Guan Y, Zheng BJ, He YQ, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. *Science*. 2003;302:276-278.
- 150. Chan PKS, Tang JW, Hui DSC. SARS: clinical presentation, transmission, pathogenesis and treatment options. *Clin Sci* (*Lond*). 2006;110:193-204.
- 151. Drosten C, Günther S, Preiser W, et al. Identification of a novel coronavirus in patients with severe acute respiratory syndrome. *N Engl J Med.* 2003;348:1967-1976.
- 152. Ksiazek TG, Erdman D, Goldsmith CS, et al. A novel coronavirus associated with severe acute respiratory syndrome. N Engl J Med. 2003;348:1953-1966.
- 153. Gralinski LE, Menachery VD, Morgan AP, et al. Allelic variation in the toll-like receptor adaptor protein Ticam2 contributes to SARS-coronavirus pathogenesis in mice. *G3 (Bethesda)*. 2017;7:1653-1663.
- Lang Z-W, Zhang L-J, Zhang S-J, et al. A clinicopathological study of three cases of severe acute respiratory syndrome (SARS). *Pathology*. 2003;35:526-531.
- Nicholls JM, Poon LLM, Lee KC, et al. Lung pathology of fatal severe acute respiratory syndrome. *Lancet*. 2003;361:1773-1778.
- Cheung OY, Chan JWM, Ng CK, Koo CK. The spectrum of pathological changes in severe acute respiratory syndrome (SARS). *Histopathology*. 2004;45:119-124.
- 157. Gu J, Korteweg C. Pathology and pathogenesis of severe acute respiratory syndrome. *Am J Pathol*. 2007;170:1136-1147.
- Hsiao CH, Wu M-Z, Chen C-L, et al. Evolution of pulmonary pathology in severe acute respiratory syndrome. *J Formos Med Assoc.* 2005;104:75-81.
- 159. Chan KS, Zheng JP, Mok YW, et al. SARS: prognosis, outcome and sequelae. *Respirology*. 2003;8(Suppl):S36-S40.
- 160. Rathinam VAK, Fitzgerald KA. Cytosolic surveillance and antiviral immunity. *Curr Opin Virol*. 2011;1:455-462.
- 161. Jensen S, Thomsen AR. Sensing of RNA viruses: a review of innate immune receptors involved in recognizing RNA virus invasion. *J Virol.* 2012;86:2900-2910.
- Binnie A, Tsang JLY, dos Santos CC. Biomarkers in acute respiratory distress syndrome. *Curr Opin Crit Care*. 2014;20:47-55.
- 163. Williams AE, Chambers RC. The mercurial nature of neutrophils: still an enigma in ARDS? Am J Physiol Lung Cell Mol Physiol. 2014;306:L217-L30.

- 164. Frieman MB, Chen J, Morrison TE, et al. SARS-CoV pathogenesis is regulated by a STAT1 dependent but a type I, II and III interferon receptor independent mechanism. *PLoS Pathog.* 2010;6:e1000849-e.
- 165. Hogan RJ, Gao G, Rowe T, et al. Resolution of primary severe acute respiratory syndrome-associated coronavirus infection requires Stat1. *J Virol.* 2004;78:11416-11421.
- 166. Totura AL, Baric RS. SARS coronavirus pathogenesis: host innate immune responses and viral antagonism of interferon. *Curr Opin Virol.* 2012;2:264-275.
- 167. Snijder EJ, van der Meer Y, Zevenhoven-Dobbe J, et al. Ultrastructure and origin of membrane vesicles associated with the severe acute respiratory syndrome coronavirus replication complex. *J Virol.* 2006;80:5927-5940.
- 168. Knoops K, Kikkert M, Worm SHEvd, et al. SARS-coronavirus replication is supported by a reticulovesicular network of modified endoplasmic reticulum. *PLoS Biol.* 2008;6: e226-e.
- 169. Chen Y, Cai H, Pan JA, et al. Functional screen reveals SARS coronavirus nonstructural protein nsp14 as a novel cap N7 methyltransferase. *Proc Natl Acad Sci U S A*. 2009;106:3484-3489.
- Bouvet M, Debarnot C, Imbert I, et al. In vitro reconstitution of SARS-coronavirus mRNA cap methylation. *PLoS Pathog.* 2010;6:e1000863-e.
- 171. Chen Y, Su C, Ke M, et al. Biochemical and structural insights into the mechanisms of SARS coronavirus RNA ribose 2'-Omethylation by nsp16/nsp10 protein complex. *PLoS Pathog.* 2011;7:e1002294-e.
- 172. Züst R, Cervantes-Barragan L, Habjan M, et al. Ribose 2'-Omethylation provides a molecular signature for the distinction of self and non-self mRNA dependent on the RNA sensor Mda5. *Nat Immunol.* 2011;12:137-143.
- 173. Wathelet MG, Orr M, Frieman MB, Baric RS. Severe acute respiratory syndrome coronavirus evades antiviral signaling: role of nsp1 and rational design of an attenuated strain. *J Virol.* 2007;81:11620-11633.
- 174. Kamitani W, Huang C, Narayanan K, Lokugamage KG, Makino S. A two-pronged strategy to suppress host protein synthesis by SARS coronavirus Nsp1 protein. *Nat Struct Mol Biol.* 2009;16:1134-1140.
- 175. Huang C, Lokugamage KG, Rozovics JM, Narayanan K, Semler BL, Makino S. SARS coronavirus nsp1 protein induces template-dependent endonucleolytic cleavage of mRNAs: viral mRNAs are resistant to nsp1-induced RNA cleavage. *PLoS Pathog.* 2011;7:e1002433-e.
- 176. Devaraj SG, Wang N, Chen Z, et al. Regulation of IRF-3-dependent innate immunity by the papain-like protease domain of the severe acute respiratory syndrome coronavirus. *J Biol Chem.* 2007;282:32208-32221.
- 177. Frieman M, Ratia K, Johnston RE, Mesecar AD, Baric RS. Severe acute respiratory syndrome coronavirus papain-like protease ubiquitin-like domain and catalytic domain regulate antagonism of IRF3 and NF-kappaB signaling. *J Virol.* 2009;83:6689-6705.
- 178. Sun L, Xing Y, Chen X, et al. Coronavirus papain-like proteases negatively regulate antiviral innate immune response through disruption of STING-mediated signaling. *PLoS One*. 2012;7:e30802-e.

- 179. Cui L, Wang H, Ji Y, et al. The nucleocapsid protein of coronaviruses acts as a viral suppressor of RNA silencing in mammalian cells. *J Virol*. 2015;89:9029-9043.
- 180. Siu K-L, Kok K-H, Ng M-HJ, et al. Severe acute respiratory syndrome coronavirus M protein inhibits type I interferon production by impeding the formation of TRAF3.TANK.TBK1/IKKepsilon complex. J Biol Chem. 2009;284:16202-16209.
- 181. Freundt EC, Yu L, Park E, Lenardo MJ, Xu X-N. Molecular determinants for subcellular localization of the severe acute respiratory syndrome coronavirus open reading frame 3b protein. *J Virol.* 2009;83:6631-6640.
- 182. Kopecky-Bromberg SA, Martínez-Sobrido L, Frieman M, Baric RA, Palese P. Severe acute respiratory syndrome coronavirus open reading frame (ORF) 3b, ORF 6, and nucleocapsid proteins function as interferon antagonists. *J Virol.* 2007;81:548-557.
- van den Brand JMA, Smits SL, Haagmans BL. Pathogenesis of Middle East respiratory syndrome coronavirus. J Pathol. 2015;235:175-184.
- Rasmussen SA, Watson AK, Swerdlow DL. Middle East respiratory syndrome (MERS). *Microbiol Spectr.* 2016;4.
- 185. Shi Z, Hu Z. A review of studies on animal reservoirs of the SARS coronavirus. *Virus Res.* 2008;133:74-87.
- 186. de Groot RJ, Baker SC, Baric RS, et al. Middle East respiratory syndrome coronavirus (MERS-CoV): announcement of the Coronavirus Study Group. *J Virol*. 2013;87:7790-7792.
- 187. Reusken CBEM, Haagmans BL, Müller MA, et al. Middle East respiratory syndrome coronavirus neutralising serum antibodies in dromedary camels: a comparative serological study. *Lancet Infect Dis.* 2013;13:859-866.
- Azhar EI, El-Kafrawy SA, Farraj SA, et al. Evidence for camelto-human transmission of MERS coronavirus. *N Engl J Med.* 2014;370:2499-2505.
- 189. Zumla A, Hui DS, Perlman S. Middle East respiratory syndrome. *Lancet*. 2015;386:995-1007.
- 190. Chan RWY, Chan MCW, Agnihothram S, et al. Tropism of and innate immune responses to the novel human betacoronavirus lineage C virus in human ex vivo respiratory organ cultures. J Virol. 2013;87:6604-6614.
- 191. Zhou J, Chu H, Li C, et al. Active replication of Middle East respiratory syndrome coronavirus and aberrant induction of inflammatory cytokines and chemokines in human macrophages: implications for pathogenesis. J Infect Dis. 2014;209:1331-1342.
- 192. Zhao J, Li K, Wohlford-Lenane C, et al. Rapid generation of a mouse model for Middle East respiratory syndrome. *Proc Natl Acad Sci U S A*. 2014;111:4970-4975.
- 193. Oudshoorn D, van der Hoeven B, Limpens RWAL, et al. Antiviral innate immune response interferes with the formation of replication-associated membrane structures induced by a positive-strand RNA virus. *mBio*. 2016;7:e01991-16.
- Menachery VD, Debbink K, Baric RS. Coronavirus nonstructural protein 16: evasion, attenuation, and possible treatments. *Virus Res.* 2014;194:191-199.
- 195. Yang Y, Zhang L, Geng H, et al. The structural and accessory proteins M, ORF 4a, ORF 4b, and ORF 5 of Middle East respiratory syndrome coronavirus (MERS-CoV) are potent interferon antagonists. *Protein Cell*. 2013;4:951-961.

- 197. Siu K-L, Yeung ML, Kok K-H, et al. Middle east respiratory syndrome coronavirus 4a protein is a double-stranded RNAbinding protein that suppresses PACT-induced activation of RIG-I and MDA5 in the innate antiviral response. *J Virol.* 2014;88:4866-4876.
- 198. Yang Y, Ye F, Zhu N, et al. Middle East respiratory syndrome coronavirus ORF4b protein inhibits type I interferon production through both cytoplasmic and nuclear targets. *Sci Rep.* 2015;5:17554.
- 199. Bailey-Elkin BA, Knaap RCM, Johnson GG, et al. Crystal structure of the Middle East respiratory syndrome coronavirus (MERS-CoV) papain-like protease bound to ubiquitin facilitates targeted disruption of deubiquitinating activity to demonstrate its role in innate immune suppression. *J Biol Chem.* 2014;289:34667-34682.
- 200. Lokugamage KG, Narayanan K, Nakagawa K, et al. Middle East respiratory syndrome coronavirus nsp1 inhibits host gene expression by selectively targeting mRNAs transcribed in the nucleus while sparing mRNAs of cytoplasmic origin. *J Virol.* 2015;89:10970-10981.
- 201. Alraddadi BM, Watson JT, Almarashi A, et al. Risk factors for primary Middle East respiratory syndrome coronavirus illness in humans, Saudi Arabia, 2014. *Emerg Infect Dis.* 2016;22:49-55.
- 202. Nam H-S, Park JW, Ki M, Yeon M-Y, Kim J, Kim SW. High fatality rates and associated factors in two hospital outbreaks of MERS in Daejeon, the Republic of Korea. *Int J Infect Dis.* 2017;58:37-42.
- 203. Evans MJ, Cabral LJ, Stephens RJ, Freeman G. Renewal of alveolar epithelium in the rat following exposure to NO2. *Am J Pathol.* 1973;70:175-198.
- 204. Dahlin K, Mager EM, Allen L, et al. Identification of genes differentially expressed in rat alveolar type I cells. *Am J Respir Cell Mol Biol.* 2004;31:309-316.
- 205. Ng DL, Al Hosani F, Keating MK, et al. Clinicopathologic, immunohistochemical, and ultrastructural findings of a fatal case of Middle East respiratory syndrome coronavirus infection in the United Arab Emirates, April 2014. *Am J Pathol.* 2016;186:652-658.
- 206. Alsaad KO, Hajeer AH, Al Balwi M, et al. Histopathology of Middle East respiratory syndrome coronovirus (MERS-CoV) infection - clinicopathological and ultrastructural study. *Histopathology*. 2018;72:516-524.
- 207. Haagmans BL, Kuiken T, Martina BE, et al. Pegylated interferon-alpha protects type 1 pneumocytes against SARS coronavirus infection in macaques. *Nat Med.* 2004;10:290-293.
- 208. Seys LJM, Widagdo W, Verhamme FM, et al. DPP4, the Middle East respiratory syndrome coronavirus receptor, is upregulated in lungs of smokers and chronic obstructive pulmonary disease patients. *Clin Infect Dis.* 2018;66:45-53.
- 209. Holshue ML, DeBolt C, Lindquist S, et al. First Case of 2019 Novel Coronavirus in the United States. N Engl J Med. 2020;382(10):929-936.
- 210. Zhu N, Zhang D, Wang W, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med.* 2020;382:727-733.
- 211. Chan JF-W, Kok K-H, Zhu Z, et al. Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from

a patient with atypical pneumonia after visiting Wuhan. *Emerg Microbes Infect.* 2020;9:221-236.

- 212. Hu D, Zhu C, Ai L, et al. Genomic characterization and infectivity of a novel SARS-like coronavirus in Chinese bats. *Emerg Microbes Infect.* 2018;7:154.
- 213. Chan JF-W, Yuan S, Kok K-H, et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster. *Lancet.* 2020;395:514-523.
- 214. Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet*. 2020;395:507-513.
- 215. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan. *China Lancet*. 2020;395:497-506.
- Zhou P, Yang X-L, Wang X-G, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature*. 2020;579(7798):270-273.
- 217. Assiri A, Al-Tawfiq JA, Al-Rabeeah AA, et al. Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. *Lancet Infect Dis.* 2013;13:752-761.
- Liu W, Morse JS, Lalonde T, Xu S. Learning from the past: possible urgent prevention and treatment options for severe acute respiratory infections caused by 2019-nCoV. *Chembiochem*. 2020;21(5):730-738.
- 219. Guan GW, Gao L, Wang JW, et al. Exploring the mechanism of liver enzyme abnormalities in patients with novel coronavirus-infected pneumonia. *Zhonghua Gan Zang Bing Za Zhi*. 2020;28:E002-E.
- 220. Tian HY. 2019-nCoV: new challenges from coronavirus. *Zhonghua Yu Fang Yi Xue Za Zhi*. 2020;54:E001-E.
- 221. Xu Z, Shi L, Wang Y, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med.* 2020;8(4):420-422.
- 222. Tan L, Wang Q, Zhang D, et al. Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. *Signal Transduct Target Ther.* 2020;5:33.
- 223. Habibzadeh P, Stoneman EK. The Novel Coronavirus: a Bird's Eye View. *Int J Occup Environ Med*. 2020;11:65-71.
- 224. Cheng VCC, Tang BSF, Wu AKL, Chu CM, Yuen KY. Medical treatment of viral pneumonia including SARS in immunocompetent adult. *J Infect*. 2004;49:262-273.
- 225. Cheng VCC, Chan JFW, To KKW, Yuen KY. Clinical management and infection control of SARS: lessons learned. *Antiviral Res.* 2013;100:407-419.
- 226. Zumla A, Chan JFW, Azhar EI, Hui DSC, Yuen K-Y. Coronaviruses - drug discovery and therapeutic options. *Nat Rev Drug Discov*. 2016;15:327-347.
- 227. Morse JS, Lalonde T, Xu S, Liu WR. Learning from the past: possible urgent prevention and treatment options for severe acute respiratory infections caused by 2019-nCoV. *Chembiochem.* 2020;21(5):730-738.
- 228. Lu L, Liu Q, Zhu Y, et al. Structure-based discovery of Middle East respiratory syndrome coronavirus fusion inhibitor. *Nat Commun.* 2014;5:3067.
- 229. He Y, Li J, Li W, Lustigman S, Farzan M, Jiang S. Crossneutralization of human and palm civet severe acute respiratory syndrome coronaviruses by antibodies targeting

the receptor-binding domain of spike protein. *J Immunol.* 2006;176:6085-6092.

- 230. Yuan K, Yi L, Chen J, et al. Suppression of SARS-CoV entry by peptides corresponding to heptad regions on spike glycoprotein. *Biochem Biophys Res Commun.* 2004;319:746-752.
- 231. Gao J, Lu G, Qi J, et al. Structure of the fusion core and inhibition of fusion by a heptad repeat peptide derived from the S protein of Middle East respiratory syndrome coronavirus. *J Virol.* 2013;87:13134-13140.
- 232. Jiang L, Wang N, Zuo T, et al. Potent neutralization of MERS-CoV by human neutralizing monoclonal antibodies to the viral spike glycoprotein. *Sci Transl Med.* 2014;6:234ra59-ra59.
- 233. Tang X-C, Agnihothram SS, Jiao Y, et al. Identification of human neutralizing antibodies against MERS-CoV and their role in virus adaptive evolution. *Proc Natl Acad Sci U S A*. 2014;111:E2018-E26.
- 234. Ying T, Du L, Ju TW, et al. Exceptionally potent neutralization of Middle East respiratory syndrome coronavirus by human monoclonal antibodies. *J Virol.* 2014;88:7796-7805.
- 235. Shirato K, Kawase M, Matsuyama S. Middle East respiratory syndrome coronavirus infection mediated by the transmembrane serine protease TMPRSS2. *J Virol.* 2013;87:12552-12561.
- 236. Qian Z, Dominguez SR, Holmes KV. Role of the spike glycoprotein of human Middle East respiratory syndrome coronavirus (MERS-CoV) in virus entry and syncytia formation. *PLoS One.* 2013;8:e76469-e.
- 237. Zhou Y, Vedantham P, Lu K, et al. Protease inhibitors targeting coronavirus and filovirus entry. *Antiviral Res.* 2015;116:76-84.
- 238. van Boheemen S, de Graaf M, Lauber C, et al. Genomic characterization of a newly discovered coronavirus associated with acute respiratory distress syndrome in humans. *mBio*. 2012;3:e00473-12.
- 239. Lundin A, Dijkman R, Bergström T, et al. Targeting membranebound viral RNA synthesis reveals potent inhibition of diverse coronaviruses including the Middle East respiratory syndrome virus. *PLoS Pathog.* 2014;10:e1004166-e.
- 240. Josset L, Menachery VD, Gralinski LE, et al. Cell host response to infection with novel human coronavirus EMC predicts potential antivirals and important differences with SARS coronavirus. *mBio*. 2013;4:e00165.
- 241. Lau SKP, Lau CCY, Chan K-H, et al. Delayed induction of proinflammatory cytokines and suppression of innate antiviral response by the novel Middle East respiratory syndrome coronavirus: implications for pathogenesis and treatment. *J Gen Virol.* 2013;94:2679-2690.
- 242. Hart BJ, Dyall J, Postnikova E, et al. Interferon-β and mycophenolic acid are potent inhibitors of Middle East respiratory syndrome coronavirus in cell-based assays. J Gen Virol. 2014;95:571-577.
- 243. Kindrachuk J, Ork B, Hart BJ, et al. Antiviral potential of ERK/MAPK and PI3K/AKT/mTOR signaling modulation for Middle East respiratory syndrome coronavirus infection as identified by temporal kinome analysis. *Antimicrob Agents Chemother*. 2015;59:1088-1099.
- 244. Pfefferle S, Schöpf J, Kögl M, et al. The SARS-coronavirus-host interactome: identification of cyclophilins as target for pancoronavirus inhibitors. *PLoS Pathog*. 2011;7:e1002331-e.
- 245. de Wilde AH, Raj VS, Oudshoorn D, et al. MERS-coronavirus replication induces severe in vitro cytopathology and is strongly

inhibited by cyclosporin A or interferon- α treatment. *J Gen Virol.* 2013;94:1749-1760.

- 246. Eltahla AA, Luciani F, White PA, Lloyd AR, Bull RA. Inhibitors of the hepatitis c virus polymerase; mode of action and resistance. *Viruses*. 2015;7:5206-5224.
- 247. De Clercq E, Li G. Approved antiviral drugs over the past 50 years. *Clin Microbiol Rev.* 2016;29:695-747.
- 248. De Clercq E. New nucleoside analogues for the treatment of hemorrhagic fever virus infections. *Chem Asian J.* 2019;14:3962-3968.
- 249. Peters HL, Jochmans D, de Wilde AH, et al. Design, synthesis and evaluation of a series of acyclic fleximer nucleoside analogues with anti-coronavirus activity. *Bioorg Med Chem Lett.* 2015;25:2923-2926.
- Li G, De Clercq E. Therapeutic options for the 2019 novel coronavirus (2019-nCoV). Nat Rev Drug Discov. 2020;19:149-150.
- 251. Baranovich T, Wong S-S, Armstrong J, et al. T-705 (favipiravir) induces lethal mutagenesis in influenza A H1N1 viruses in vitro. *J Virol.* 2013;87:3741-3751.
- 252. Gowen BB, Wong M-H, Jung K-H, et al. In vitro and in vivo activities of T-705 against arenavirus and bunyavirus infections. *Antimicrob Agents Chemother*. 2007;51:3168-3176.
- 253. Julander JG, Shafer K, Smee DF, Morrey JD, Furuta Y. Activity of T-705 in a hamster model of yellow fever virus infection in comparison with that of a chemically related compound, T-1106. Antimicrob Agents Chemother. 2009;53:202-209.
- 254. Smither SJ, Eastaugh LS, Steward JA, Nelson M, Lenk RP, Lever MS. Post-exposure efficacy of oral T-705 (Favipiravir) against inhalational Ebola virus infection in a mouse model. *Antiviral Res.* 2014;104:153-155.
- 255. Zaraket H, Saito R. Japanese surveillance systems and treatment for influenza. *Curr Treat Options Infect Dis.* 2016;8:311-328.
- 256. Oestereich L, Lüdtke A, Wurr S, Rieger T, Muñoz-Fontela C, Günther S. Successful treatment of advanced Ebola virus infection with T-705 (favipiravir) in a small animal model. *Antiviral Res.* 2014;105:17-21.
- 257. Sidwell RW, Barnard DL, Day CW, et al. Efficacy of orally administered T-705 on lethal avian influenza A (H5N1) virus infections in mice. *Antimicrob Agents Chemother*. 2007;51:845-851.
- Bai C-Q, Mu J-S, Kargbo D, et al. Clinical and virological characteristics of Ebola virus disease patients treated with Favipiravir (T-705)-Sierra Leone, 2014. *Clin Infect Dis.* 2016;63:1288-1294.
- 259. Wang M, Cao R, Zhang L, et al. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res.* 2020;30(3):269-271.
- Sidwell RW, Huffman JH, Khare GP, Allen LB, Witkowski JT, Robins RK. Broad-spectrum antiviral activity of Virazole: 1-beta-D-ribofuranosyl-1,2,4-triazole-3-carboxamide. *Science*. 1972;177:705-706.
- Ölschläger S, Neyts J, Günther S. Depletion of GTP pool is not the predominant mechanism by which ribavirin exerts its antiviral effect on Lassa virus. *Antiviral Res.* 2011;91: 89-93.
- 262. So LKY, Lau ACW, Yam LYC, et al. Development of a standard treatment protocol for severe acute respiratory syndrome. *Lancet*. 2003;361:1615-1617.

263. Wong SSY, Yuen K-Y. The management of coronavirus infections with particular reference to SARS. *J Antimicrob Chemother*. 2008;62:437-441.

LIN ET AL.

- 264. Falzarano D, de Wit E, Rasmussen AL, et al. Treatment with interferon-α2b and ribavirin improves outcome in MERS-CoV-infected rhesus macaques. *Nat Med.* 2013;19:1313-1317.
- 265. Omrani AS, Saad MM, Baig K, et al. Ribavirin and interferon alfa-2a for severe Middle East respiratory syndrome coronavirus infection: a retrospective cohort study. *Lancet Infect Dis.* 2014;14:1090-1095.
- 266. Al-Tawfiq JA, Momattin H, Dib J, Memish ZA. Ribavirin and interferon therapy in patients infected with the Middle East respiratory syndrome coronavirus: an observational study. *Int J Infect Dis.* 2014;20:42-46.
- 267. Shalhoub S, Farahat F, Al-Jiffri A, et al. IFN-α2a or IFN-βla in combination with ribavirin to treat Middle East respiratory syndrome coronavirus pneumonia: a retrospective study. JAntimicrob Chemother. 2015;70:2129-2132.
- 268. Hung IF, Lung KC, Tso EY, et al. Triple combination of interferon beta-1b, lopinavir-ritonavir, and ribavirin in the treatment of patients admitted to hospital with COVID-19: an open-label, randomised, phase 2 trial. *Lancet*. 2020;395:1695-1704.
- 269. Tchesnokov EP, Feng JY, Porter DP, Götte M. Mechanism of inhibition of Ebola virus RNA-dependent RNA polymerase by Remdesivir. *Viruses*. 2019;11:326.
- Warren TK, Jordan R, Lo MK, et al. Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys. *Nature*. 2016;531:381-385.
- 271. Agostini ML, Andres EL, Sims AC, et al. Coronavirus susceptibility to the antiviral Remdesivir (GS-5734) is mediated by the viral polymerase and the proofreading exoribonuclease. *mBio*. 2018;9:e00221-18.
- 272. Sheahan TP, Sims AC, Graham RL, et al. Broad-spectrum antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses. *Sci Transl Med.* 2017;9:eaal3653.
- 273. Lo MK, Jordan R, Arvey A, et al. GS-5734 and its parent nucleoside analog inhibit Filo-, Pneumo-, and Paramyxoviruses. *Sci Rep.* 2017;7:43395.
- 274. Wang Y, Zhang D, Du G, et al. Remdesivir in adults with severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. *Lancet*. 2020;395:1569-1578.
- 275. Grein J, Ohmagari N, Shin D, et al. Compassionate use of Remdesivir for patients with severe Covid-19. *N Engl J Med.* 2020;382:2327-2336.
- 276. Beigel JH, Tomashek KM, Dodd LE, et al. Remdesivir for the treatment of Covid-19 preliminary report. *N Engl J Med*. 2020.
- 277. Warren TK, Wells J, Panchal RG, et al. Protection against filovirus diseases by a novel broad-spectrum nucleoside analogue BCX4430. *Nature*. 2014;508:402-405.
- 278. Delang L, Segura Guerrero N, Tas A, et al. Mutations in the chikungunya virus non-structural proteins cause resistance to favipiravir (T-705), a broad-spectrum antiviral. *J Antimicrob Chemother*. 2014;69:2770-2784.
- 279. Barretto N, Jukneliene D, Ratia K, Chen Z, Mesecar AD, Baker SC. The papain-like protease of severe acute respiratory syndrome coronavirus has deubiquitinating activity. *J Virol.* 2005;79:15189-15198.
- Mielech AM, Chen Y, Mesecar AD, Baker SC. Nidovirus papain-like proteases: multifunctional enzymes with pro-

tease, deubiquitinating and deISGylating activities. *Virus Res.* 2014;194:184-190.

- 281. Mielech AM, Kilianski A, Baez-Santos YM, Mesecar AD, Baker SC. MERS-CoV papain-like protease has deISGylating and deubiquitinating activities. *Virology*. 2014;450-451:64-70.
- 282. Cheng K-W, Cheng S-C, Chen W-Y, et al. Thiopurine analogs and mycophenolic acid synergistically inhibit the papain-like protease of Middle East respiratory syndrome coronavirus. *Antiviral Res.* 2015;115:9-16.
- 283. Lee H, Ren J, Pesavento RP, et al. Identification and design of novel small molecule inhibitors against MERS-CoV papainlike protease via high-throughput screening and molecular modeling. *Bioorg Med Chem.* 2019;27:1981-1989.
- Báez-Santos YM, St John SE, Mesecar AD. The SARScoronavirus papain-like protease: structure, function and inhibition by designed antiviral compounds. *Antiviral Res.* 2015;115:21-38.
- 285. Ratia K, Pegan S, Takayama J, et al. A noncovalent class of papain-like protease/deubiquitinase inhibitors blocks SARS virus replication. *Proc Natl Acad Sci U S A*. 2008;105:16119-16124.
- Lin M-H, Moses DC, Hsieh C-H, et al. Disulfiram can inhibit MERS and SARS coronavirus papain-like proteases via different modes. *Antiviral Res.* 2018;150:155-163.
- 287. Chan KS, Lai ST, Chu CM, et al. Treatment of severe acute respiratory syndrome with lopinavir/ritonavir: a multicentre retrospective matched cohort study. *Hong Kong Med J.* 2003;9:399-406.
- 288. Chu CM, Cheng VCC, Hung IFN, et al. Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. *Thorax*. 2004;59:252-256.
- 289. Chan JFW, Chan K-H, Kao RYT, et al. Broad-spectrum antivirals for the emerging Middle East respiratory syndrome coronavirus. *J Infect*. 2013;67:606-616.
- 290. Chan JF-W, Yao Y, Yeung M-L, et al. Treatment with lopinavir/ritonavir or interferon-βlb improves outcome of MERS-CoV infection in a nonhuman primate model of common marmoset. *J Infect Dis.* 2015;212:1904-1913.
- 291. Cao B, Wang Y, Wen D, et al. A trial of lopinavir-ritonavir in adults hospitalized with severe Covid-19. *N Engl J Med.* 2020;382:1787-1799.
- 292. Kim MK, Yu M-S, Park HR, et al. 2,6-Bis-arylmethyloxy-5hydroxychromones with antiviral activity against both hepatitis C virus (HCV) and SARS-associated coronavirus (SCV). *Eur J Med Chem*. 2011;46:5698-5704.
- 293. Zaher NH, Mostafa MI, Altaher AY. Design, synthesis and molecular docking of novel triazole derivatives as potential CoV helicase inhibitors. *Acta Pharm.* 2020;70:145-159.
- 294. Adedeji AO, Singh K, Kassim A, et al. Evaluation of SSYA10-001 as a replication inhibitor of severe acute respiratory syndrome, mouse hepatitis, and Middle East respiratory syndrome coronaviruses. *Antimicrob Agents Chemother*. 2014;58:4894-4898.
- 295. O'Keefe BR, Giomarelli B, Barnard DL, et al. Broad-spectrum in vitro activity and in vivo efficacy of the antiviral protein griffithsin against emerging viruses of the family Coronaviridae. *J Virol.* 2010;84:2511-2521.
- 296. Barton C, Kouokam JC, Lasnik AB, et al. Activity of and effect of subcutaneous treatment with the broad-spectrum antiviral

lectin griffithsin in two laboratory rodent models. *Antimicrob Agents Chemother*. 2014;58:120-127.

- 297. Sainz B, Jr, Mossel EC, Gallaher WR, et al. Inhibition of severe acute respiratory syndrome-associated coronavirus (SARS-CoV) infectivity by peptides analogous to the viral spike protein. *Virus Res.* 2006;120:146-155.
- 298. Xia S, Xu W, Wang Q, et al. Peptide-based membrane fusion inhibitors targeting HCoV-229E spike protein HR1 and HR2 domains. *Int J Mol Sci.* 2018;19:487.
- 299. Channappanavar R, Lu L, Xia S, et al. Protective effect of intranasal regimens containing peptidic middle east respiratory syndrome coronavirus fusion inhibitor against MERS-CoV infection. J Infect Dis. 2015;212:1894-1903.
- 300. Sun Y, Zhang H, Shi J, Zhang Z, Gong R. Identification of a novel inhibitor against middle east respiratory syndrome coronavirus. *Viruses*. 2017;9:255.
- 301. Li B-J, Tang Q, Cheng D, et al. Using siRNA in prophylactic and therapeutic regimens against SARS coronavirus in Rhesus macaque. *Nat Med.* 2005;11:944-951.
- Wu C-J, Huang H-W, Liu C-Y, Hong C-F, Chan Y-L. Inhibition of SARS-CoV replication by siRNA. *Antiviral Res.* 2005;65:45-48.
- 303. Zhang Y, Li T, Fu L, et al. Silencing SARS-CoV spike protein expression in cultured cells by RNA interference. *FEBS Lett.* 2004;560:141-146.
- 304. Akerström S, Mirazimi A, Tan Y-J. Inhibition of SARS-CoV replication cycle by small interference RNAs silencing specific SARS proteins, 7a/7b, 3a/3b and S. *Antiviral Res.* 2007;73:219-227.
- 305. He ML, Zheng BJ, Chen Y, et al. Development of interfering RNA agents to inhibit SARS-associated coronavirus infection and replication. *Hong Kong Med J.* 2009;15:28-31.
- 306. Faith SA, Sweet TJ, Bailey E, Booth T, Docherty JJ. Resveratrol suppresses nuclear factor-kappaB in herpes simplex virus infected cells. *Antiviral Res.* 2006;72:242-251.
- 307. Kimbrough CW, Lakshmanan J, Matheson PJ, et al. Resveratrol decreases nitric oxide production by hepatocytes during inflammation. *Surgery*. 2015;158:1095-1101.
- 308. Lin C-J, Lin H-J, Chen T-H, et al. Polygonum cuspidatum and its active components inhibit replication of the influenza virus through toll-like receptor 9-induced interferon beta expression. *PLoS One.* 2015;10:e0117602-e.
- 309. Francioso A, Cossi R, Fanelli S, Mastromarino P, Mosca L. Studies on trans-resveratrol/carboxymethylated (1,3/1,6)-β-dglucan association for aerosol pharmaceutical applications. *Int J Mol Sci.* 2017;18:967.
- 310. Wang L-L, Shi D-L, Gu H-Y, et al. Resveratrol attenuates inflammatory hyperalgesia by inhibiting glial activation in mice spinal cords. *Mol Med Rep.* 2016;13:4051-4057.
- 311. Zang N, Xie X, Deng Y, et al. Resveratrol-mediated gamma interferon reduction prevents airway inflammation and airway hyperresponsiveness in respiratory syncytial virus-infected immunocompromised mice. *J Virol.* 2011;85:13061-13068.
- Lin S-C, Ho C-T, Chuo W-H, Li S, Wang TT, Lin C-C. Effective inhibition of MERS-CoV infection by resveratrol. *BMC Infect Dis.* 2017;17:144.
- 313. Wilson L, Gage P, Ewart G. Hexamethylene amiloride blocks E protein ion channels and inhibits coronavirus replication. *Virology*. 2006;353:294-306.

- Dyall J, Coleman CM, Hart BJ, et al. Repurposing of clinically developed drugs for treatment of Middle East respiratory syndrome coronavirus infection. *Antimicrob Agents Chemother*. 2014;58:4885-4893.
- 315. Hollmann A, Castanho MARB, Lee B, Santos NC. Singlet oxygen effects on lipid membranes: implications for the mechanism of action of broad-spectrum viral fusion inhibitors. *Biochem J.* 2014;459:161-170.
- Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Treatment of SARS with human interferons. *Lancet*. 2003;362:293-294.
- 317. Falzarano D, de Wit E, Martellaro C, Callison J, Munster VJ, Feldmann H. Inhibition of novel β coronavirus replication by a combination of interferon- α 2b and ribavirin. *Sci Rep.* 2013;3:1686.
- 318. Morgenstern B, Michaelis M, Baer PC, Doerr HW. Ribavirin and interferon-beta synergistically inhibit SARS-associated coronavirus replication in animal and human cell lines. *Biochem Biophys Res Commun.* 2005;326:905-908.
- 319. Rossignol J-F. Nitazoxanide: a first-in-class broad-spectrum antiviral agent. *Antiviral Res.* 2014;110:94-103.
- 320. Haffizulla J, Hartman A, Hoppers M, et al. Effect of nitazoxanide in adults and adolescents with acute uncomplicated influenza: a double-blind, randomised, placebo-controlled, phase 2b/3 trial. *Lancet Infect Dis.* 2014;14:609-618.
- 321. Rossignol JF, Kabil SM, El-Gohary Y, Elfert A, Keeffe EB. Clinical trial: randomized, double-blind, placebo-controlled study of nitazoxanide monotherapy for the treatment of patients with chronic hepatitis C genotype 4. *Aliment Pharmacol Ther*. 2008;28:574-580.
- 322. Okada H, Kalinski P, Ueda R, et al. Induction of CD8+ Tcell responses against novel glioma-associated antigen peptides and clinical activity by vaccinations with {alpha}-type 1 polarized dendritic cells and polyinosinic-polycytidylic acid stabilized by lysine and carboxymethylcellulose in patients with recurrent malignant glioma. J Clin Oncol. 2011;29:330-336.
- 323. Rosenfeld MR, Chamberlain MC, Grossman SA, et al. A multiinstitution phase II study of poly-ICLC and radiotherapy with concurrent and adjuvant temozolomide in adults with newly diagnosed glioblastoma. *Neuro Oncol.* 2010;12:1071-1077.
- 324. Huentelman MJ, Zubcevic J, Hernández Prada JA, et al. Structure-based discovery of a novel angiotensin-converting enzyme 2 inhibitor. *Hypertension*. 2004;44:903-906.
- 325. Han DP, Penn-Nicholson A, Cho MW. Identification of critical determinants on ACE2 for SARS-CoV entry and development of a potent entry inhibitor. *Virology*. 2006;350:15-25.
- 326. Ohnuma K, Haagmans BL, Hatano R, et al. Inhibition of Middle East respiratory syndrome coronavirus infection by anti-CD26 monoclonal antibody. *J Virol.* 2013;87:13892-13899.
- 327. Ndao M, Nath-Chowdhury M, Sajid M, et al. A cysteine protease inhibitor rescues mice from a lethal Cryptosporidium parvum infection. *Antimicrob Agents Chemother*. 2013;57:6063-6073.
- 328. Vermeire JJ, Lantz LD, Caffrey CR. Cure of hookworm infection with a cysteine protease inhibitor. *PLoS Negl Trop Dis.* 2012;6:e1680-e.
- 329. Sai JK, Suyama M, Kubokawa Y, Matsumura Y, Inami K, Watanabe S. Efficacy of camostat mesilate against dyspepsia

MedComm

- Talukdar R, Tandon RK. Pancreatic stellate cells: new target in the treatment of chronic pancreatitis. *J Gastroenterol Hepatol*. 2008;23:34-41.
- 331. de Wilde AH, Jochmans D, Posthuma CC, et al. Screening of an FDA-approved compound library identifies four small-molecule inhibitors of Middle East respiratory syndrome coronavirus replication in cell culture. *Antimicrob Agents Chemother*. 2014;58:4875-4884.
- 332. Burkard C, Verheije MH, Haagmans BL, et al. ATP1A1mediated Src signaling inhibits coronavirus entry into host cells. *J Virol.* 2015;89:4434-4448.
- 333. Yan Y, Zou Z, Sun Y, Li X, et al. Anti-malaria drug chloroquine is highly effective in treating avian influenza A H5N1 virus infection in an animal model. *Cell Research*. 2013;23:2:300–302.
- 334. Keyaerts E, Vijgen L, Maes P, Neyts J, Van Ranst M. In vitro inhibition of severe acute respiratory syndrome coronavirus by chloroquine. *Biochem Biophys Res Commun.* 2004;323:264-268.
- 335. Kono M, Tatsumi K, Imai AM, Saito K, Kuriyama T, Shirasawa H. Inhibition of human coronavirus 229E infection in human epithelial lung cells (L132) by chloroquine: involvement of p38 MAPK and ERK. *Antiviral Res.* 2008;77:150-152.
- 336. Madrid PB, Chopra S, Manger ID, et al. A systematic screen of FDA-approved drugs for inhibitors of biological threat agents. *PLoS One.* 2013;8:e60579-e.
- 337. Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases?. *Lancet Infect Dis.* 2003;3:722-727.
- 338. Vincent MJ, Bergeron E, Benjannet S, et al. Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. *Virol J.* 2005;2:69.
- 339. Borba MGS, Val FFA, Sampaio VS, et al. Effect of high vs low doses of chloroquine diphosphate as adjunctive therapy for patients hospitalized with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection: a randomized clinical trial. *JAMA Network Open.* 2020;3:e208857.
- 340. Tang W, Cao Z, Han M, et al. Hydroxychloroquine in patients with mainly mild to moderate coronavirus disease 2019: open label, randomised controlled trial. *BMJ*. 2020;369:m1849.
- 341. Gautret P, Lagier JC, Parola P, et al. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an openlabel non-randomized clinical trial. *Int J Antimicrob Agents*. 2020:105949.
- 342. Mahévas M, Tran VT, Roumier M, et al. Clinical efficacy of hydroxychloroquine in patients with covid-19 pneumonia who require oxygen: observational comparative study using routine care data. *BMJ*. 2020;369:m1844.
- 343. Geleris J, Sun Y, Platt J, et al. Observational study of hydroxychloroquine in hospitalized patients with Covid-19. N Engl J Med. 2020;382:2411-2418.
- 344. Tanaka Y, Sato Y, Sasaki T. Suppression of coronavirus replication by cyclophilin inhibitors. *Viruses*. 2013;5:1250-1260.
- 345. Carbajo-Lozoya J, Ma-Lauer Y, Malešević M, et al. Human coronavirus NL63 replication is cyclophilin A-dependent and inhibited by non-immunosuppressive cyclosporine A-derivatives including Alisporivir. *Virus Res.* 2014;184: 44-53.

- 346. Kim Y, Lee C. Porcine epidemic diarrhea virus induces caspaseindependent apoptosis through activation of mitochondrial apoptosis-inducing factor. *Virology*. 2014;460-461:180-193.
- 347. de Wilde AH, Falzarano D, Zevenhoven-Dobbe JC, et al. Alisporivir inhibits MERS- and SARS-coronavirus replication in cell culture, but not SARS-coronavirus infection in a mouse model. *Virus Res.* 2017;228:7-13.
- 348. Li HS, Kuok DIT, Cheung MC, et al. Effect of interferon alpha and cyclosporine treatment separately and in combination on Middle East Respiratory Syndrome Coronavirus (MERS-CoV) replication in a human in-vitro and ex-vivo culture model. *Antiviral Res.* 2018;155:89-96.
- 349. Coleman CM, Sisk JM, Mingo RM, Nelson EA, White JM, Frieman MB. Abelson kinase inhibitors are potent inhibitors of severe acute respiratory syndrome coronavirus and Middle East respiratory syndrome coronavirus fusion. *J Virol.* 2016;90:8924-8933.
- 350. Lee DT, Wing YK, Leung HC, et al. Factors associated with psychosis among patients with severe acute respiratory syndrome: a case-control study. *Clin Infect Dis.* 2004;39:1247-1249.
- 351. Arabi YM, Mandourah Y, Al-Hameed F, et al. Corticosteroid therapy for critically Ill patients with Middle East respiratory syndrome. *Am J Respir Crit Care Med.* 2018;197:757-767.
- Johnson RM, Vinetz JM. Dexamethasone in the management of covid -19. *BMJ*. 2020;370:m2648.
- 353. Mahase E. Covid-19: demand for dexamethasone surges as RECOVERY trial publishes preprint. *BMJ*. 2020;369:m2512.
- 354. Wu C, Chen X, Cai Y, et al. Risk factors associated with acute respiratory distress syndrome and death in patients with coronavirus disease 2019 pneumonia in Wuhan. *China JAMA Internal Med.* 2020;180:1-11.
- 355. Wang Y, Jiang W, He Q, et al. A retrospective cohort study of methylprednisolone therapy in severe patients with COVID-19 pneumonia. *Signal Transduction and Targeted Therapy*. 2020;5:57.
- 356. He Y, Zhou Y, Wu H, et al. Identification of immunodominant sites on the spike protein of severe acute respiratory syndrome (SARS) coronavirus: implication for developing SARS diagnostics and vaccines. *J Immunol*. 2004;173:4050-4057.
- 357. Spanakis N, Tsiodras S, Haagmans BL, et al. Virological and serological analysis of a recent Middle East respiratory syndrome coronavirus infection case on a triple combination antiviral regimen. *Int J Antimicrob Agents*. 2014;44:528-532.
- 358. Yeh K-M, Chiueh T-S, Siu LK, et al. Experience of using convalescent plasma for severe acute respiratory syndrome among healthcare workers in a Taiwan hospital. *J Antimicrob Chemother*. 2005;56:919-922.
- 359. Frame JD, Verbrugge GP, Gill RG, Pinneo L. The use of Lassa fever convalescent plasma in Nigeria. *Trans R Soc Trop Med Hyg.* 1984;78:319-324.
- 360. Mupapa K, Massamba M, Kibadi K, et al. Treatment of Ebola hemorrhagic fever with blood transfusions from convalescent patients. International Scientific and Technical Committee. J Infect Dis. 1999;179(Suppl 1):S18-S23.
- 361. Ruggiero HA, Pérez Isquierdo F, Milani HA, et al. Treatment of Argentine hemorrhagic fever with convalescent's plasma. 4433 cases. *Presse Med.* 1986;15:2239-2242.
- 362. Stinebaugh BJ, Schloeder FX, Johnson KM, Mackenzie RB, Entwisle G, De Alba E. Bolivian hemorrhagic fever. A report of four cases. *Am J Med.* 1966;40:217-230.

- 363. Soo YOY, Cheng Y, Wong R, et al. Retrospective comparison of convalescent plasma with continuing high-dose methyl-prednisolone treatment in SARS patients. *Clin Microbiol Infect*. 2004;10:676-678.
- 364. Cheng Y, Wong R, Soo YOY, et al. Use of convalescent plasma therapy in SARS patients in Hong Kong. Eur J Clin Microbiol Infect Dis. 2005;24:44-46.
- 365. Singh SK. Middle East respiratory syndrome virus pathogenesis. *Semin Respir Crit Care Med*. 2016;37:572-577.
- 366. Coughlin MM, Prabhakar BS. Neutralizing human monoclonal antibodies to severe acute respiratory syndrome coronavirus: target, mechanism of action, and therapeutic potential. *Rev Med Virol.* 2012;22:2-17.
- 367. Corti D, Zhao J, Pedotti M, et al. Prophylactic and postexposure efficacy of a potent human monoclonal antibody against MERS coronavirus. *Proc Natl Acad Sci U S A*. 2015;112:10473-10478.
- 368. Du L, Zhao G, Yang Y, et al. A conformation-dependent neutralizing monoclonal antibody specifically targeting receptorbinding domain in Middle East respiratory syndrome coronavirus spike protein. *J Virol.* 2014;88:7045-7053.
- 369. Li Y, Wan Y, Liu P, et al. A humanized neutralizing antibody against MERS-CoV targeting the receptor-binding domain of the spike protein. *Cell Res.* 2015;25:1237-1249.
- 370. Pascal KE, Coleman CM, Mujica AO, et al. Pre- and postexposure efficacy of fully human antibodies against spike protein in a novel humanized mouse model of MERS-CoV infection. *Proc Natl Acad Sci U S A*. 2015;112:8738-8743.
- Graham RL, Donaldson EF, Baric RS. A decade after SARS: strategies for controlling emerging coronaviruses. *Nat Rev Microbiol.* 2013;11:836-848.
- 372. Roper RL, Rehm KE. SARS vaccines: where are we? *Expert Rev Vaccines*. 2009;8:887-898.
- 373. He Y, Zhou Y, Siddiqui P, Jiang S. Inactivated SARS-CoV vaccine elicits high titers of spike protein-specific antibodies that block receptor binding and virus entry. *Biochem Biophys Res Commun.* 2004;325:445-452.
- 374. Takasuka N, Fujii H, Takahashi Y, et al. A subcutaneously injected UV-inactivated SARS coronavirus vaccine elicits systemic humoral immunity in mice. *Int Immunol.* 2004;16:1423-1430.
- 375. Enjuanes L, Dediego ML, Alvarez E, Deming D, Sheahan T, Baric R. Vaccines to prevent severe acute respiratory syndrome coronavirus-induced disease. *Virus Res.* 2008;133:45-62.
- 376. Rockx B, Donaldson E, Frieman M, et al. Escape from human monoclonal antibody neutralization affects in vitro and in vivo fitness of severe acute respiratory syndrome coronavirus. J Infect Dis. 2010;201:946-955.

- 377. Müller MA, Meyer B, Corman VM, et al. Presence of Middle East respiratory syndrome coronavirus antibodies in Saudi Arabia: a nationwide, cross-sectional, serological study. *Lancet Infect Dis.* 2015;15:559-564.
- 378. Kim Y, Liu H, Galasiti Kankanamalage AC, et al. Reversal of the progression of fatal coronavirus infection in cats by a broad-spectrum coronavirus protease inhibitor. *PLoS Pathog.* 2016;12(3):e1005531-e1005531.
- 379. Galasiti Kankanamalage AC, Kim Y, Damalanka VC, et al. Structure-guided design of potent and permeable inhibitors of MERS coronavirus 3CL protease that utilize a piperidine moiety as a novel design element. *Eur J Med Chem.* 2018;150:334-346.
- 380. Konno H, Onuma T, Nitanai I, et al. Synthesis and evaluation of phenylisoserine derivatives for the SARS-CoV 3CL protease inhibitor. *Bioorg Med Chem Lett.* 2017;27(12):2746-2751.
- Kumar V, Tan K-P, Wang Y-M, Lin S-W, Liang P-H. Identification, synthesis and evaluation of SARS-CoV and MERS-CoV 3C-like protease inhibitors. *Bioorg Med Chem*. 2016;24(13):3035-3042.
- 382. Wang L, Xu J, Kong Y, et al. Engineering a novel antibodypeptide bispecific fusion protein against MERS-CoV. Antibodies (Basel). 2019;8(4):53.
- 383. Zhao P, Wang B, Ji C-M, Cong X, Wang M, Huang Y-W. Identification of a peptide derived from the heptad repeat 2 region of the porcine epidemic diarrhea virus (PEDV) spike glycoprotein that is capable of suppressing PEDV entry and inducing neutralizing antibodies. *Antiviral Res.* 2018;150:1-8.
- 384. Zhao H, Zhou J, Zhang K, et al. A novel peptide with potent and broad-spectrum antiviral activities against multiple respiratory viruses. *Sci Rep.* 2016;6:22008-22008.
- 385. Rappe JCF, de Wilde A, Di H, et al. Antiviral activity of K22 against members of the order Nidovirales. *Virus Res.* 2018;246:28-34.
- Coleman CM, Liu YV, Mu H, et al. Purified coronavirus spike protein nanoparticles induce coronavirus neutralizing antibodies in mice. *Vaccine*. 2014;32(26):3169-3174.

How to cite this article: Lin P, Wang M, Wei Y, Kim T, Wei X. Coronavirus in human diseases: Mechanisms and advances in clinical treatment. *MedComm.* 2020;1-32. https://doi.org/10.1002/mco2.26